



US009134298B2

(12) **United States Patent**
Das Gupta et al.

(10) **Patent No.:** **US 9,134,298 B2**
(45) **Date of Patent:** ***Sep. 15, 2015**

(54) **COMPOSITIONS AND METHODS TO PREVENT CANCER WITH CUPREDOXINS**

(75) Inventors: **Tapas Das Gupta**, River Forest, IL (US); **Ananda Chakrabarty**, Villa Park, IL (US)

(73) Assignee: **The Board of Trustees of the University of Illinois**, Urbana, IL (US)

(*) Notice: Subject to any disclaimer, the term of this patent is extended or adjusted under 35 U.S.C. 154(b) by 0 days.

This patent is subject to a terminal disclaimer.

(21) Appl. No.: **13/555,376**

(22) Filed: **Jul. 23, 2012**

(65) **Prior Publication Data**

US 2012/0302509 A1 Nov. 29, 2012

Related U.S. Application Data

(63) Continuation of application No. 12/617,841, filed on Nov. 13, 2009, now Pat. No. 8,227,402, which is a continuation of application No. 11/854,654, filed on Sep. 13, 2007, now Pat. No. 7,618,939, and a continuation-in-part of application No. 11/244,105, filed on Oct. 6, 2005, now Pat. No. 7,691,383, and a continuation-in-part of application No. 10/720,603, filed on Nov. 24, 2003, now Pat. No. 7,491,394, and a continuation-in-part of application No. 10/047,710, filed on Jan. 15, 2002, now Pat. No. 7,084,105.

(60) Provisional application No. 60/844,358, filed on Sep. 14, 2006, provisional application No. 60/616,782, filed on Oct. 7, 2004, provisional application No. 60/680,500, filed on May 13, 2005, provisional application No. 60/414,550, filed on Aug. 15, 2003, provisional application No. 60/269,133, filed on Feb. 15, 2001.

(51) **Int. Cl.**

G01N 33/50 (2006.01)
C07K 14/195 (2006.01)
C07K 14/21 (2006.01)
C07K 14/22 (2006.01)
C07K 14/225 (2006.01)
C07K 14/235 (2006.01)
C07K 14/28 (2006.01)
A61K 38/16 (2006.01)

(52) **U.S. Cl.**

CPC **G01N 33/5017** (2013.01); **A61K 38/164** (2013.01); **A61K 38/168** (2013.01); **C07K 14/195** (2013.01); **C07K 14/21** (2013.01); **C07K 14/22** (2013.01); **C07K 14/225** (2013.01); **C07K 14/235** (2013.01); **C07K 14/28** (2013.01)

(58) **Field of Classification Search**

CPC **A61K 38/164**; **C07K 14/21**
USPC **514/1.1**; **530/350**
See application file for complete search history.

(56) **References Cited**

U.S. PATENT DOCUMENTS

4,897,268 A	1/1990	Tice et al.
5,075,109 A	12/1991	Tice et al.
5,811,128 A	9/1998	Tice et al.
5,814,344 A	9/1998	Tice et al.
5,820,883 A	10/1998	Tice et al.
5,853,763 A	12/1998	Tice et al.
5,928,647 A	7/1999	Rock
5,942,252 A	8/1999	Tice et al.
7,084,105 B2	8/2006	Chakrabarty et al.
7,301,010 B2	11/2007	Chakrabarty et al.
7,338,766 B2	3/2008	Chakrabarty et al.
7,381,701 B2	6/2008	Chakrabarty et al.
7,618,939 B2 *	11/2009	Gupta et al. 514/1.1
7,691,383 B2 *	4/2010	Chakrabarty et al. 424/185.1
2006/0040269 A1	2/2006	Chakrabarty et al.
2006/0149037 A1	7/2006	Chakrabarty et al.

FOREIGN PATENT DOCUMENTS

WO 2005018662 3/2005

OTHER PUBLICATIONS

Ambler 1971; Recent developments in the chemical study of protein structures. INSERM, Paris, pp. 289-305.*

Ambler et al. 1989; Two distinct azurins function in the electron-transport chain of the obligate methylotroph *Methylobacter*. *Biochem. J.* 261: 495-499.*

Hoitink et al. 1990; Isolation and sequencing of the *Alcaligenes denitrificans* azurin-encoding gene: comparison with genes encoding blue copper proteins from *Pseudomonas aeruginosa* and *Alcaligenes faecalis*. *Gene*, 90: 15-20.*

Karlsson et al. 1989; Expression of the blue copper protein azurin from *Pseudomonas aeruginosa* in *Escherichia coli*. *FEB* 246(1,2): 211-217.*

Lowenadler et al. 1986; Production of specific antibodies against protein A fusion proteins. *EMBO J.* 5 (9): 2393-2398.*

Yamada et al., *PNAS* 99(22): 14098-14103 (2002).

(Continued)

Primary Examiner — Karen Cochrane Carlson

(74) *Attorney, Agent, or Firm* — Don J. Pelto, Esq.; Sheppard, Mullin, Richter & Hampton LLP

(57)

ABSTRACT

The present invention relates to compositions comprising peptides that may be variants, derivatives and structural equivalents of cupredoxins that inhibit the development of premalignant lesions in mammalian cells, tissues and animals. Specifically, these compositions may comprise azurin from *Pseudomonas aeruginosa*, and/or the 50-77 residue region of azurin (p28). The present invention further relates to compositions that may comprise cupredoxin(s), and/or variants, derivatives or structural equivalents of cupredoxins, that retain the ability to inhibit the development of premalignant lesions in mammalian cells, tissues or animals. These compositions may be peptides or pharmaceutical compositions, among others. The compositions of the invention may be used to prevent the development of premalignant lesions in mammalian cells, tissues and animals, and thus prevent cancer.

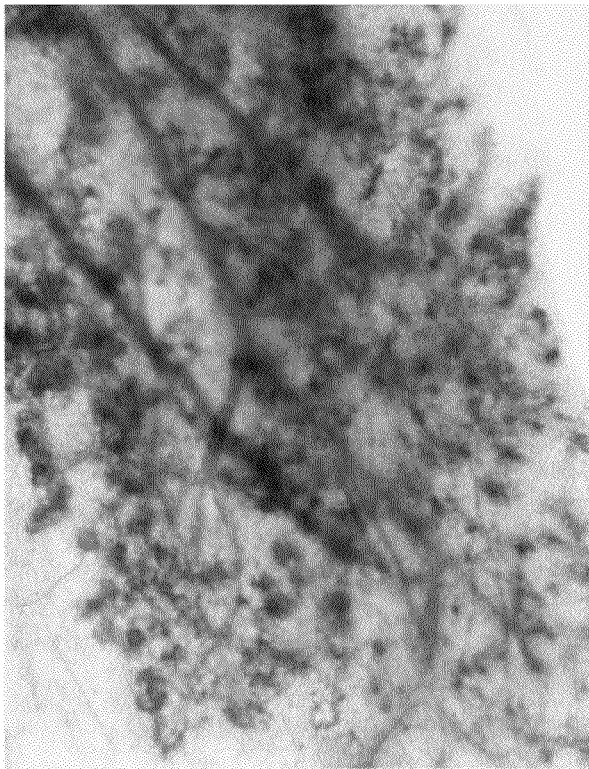
(56)

References Cited**OTHER PUBLICATIONS**

- Yamada et al., *Infect Immun.* 70: 7054-7062, (2002).
 Yamada et al., *Cell. Microbial.* 7: 1418-1431 (2005).
 Yamada et al., *Cell Cycle* 3: 1182-1187 (2004).
 Punj et al., *Oncogene.* 23: 2367-2378 (2004).
 Jang et al., *Science* 275: 218-220 (1997).
 Metha, *Eur. J. Cancer* 36: 1275-1282 (2000).
 Mehta et al., *J. Natl. Cancer Inst.* 89: 212-219 (1997).
 Mehta et al., *J. Natl. Cancer Inst.* 93: 1103-1106(2001).
 Mehta et al., *Meth Cell Sci* 19: 19-24(1997).
 Ghadiri & Fernholz, *J. Am. Chem. Soc.*, 112: 9633-9635 (1990).
 Zaborina et al. *Microbiology* 146: 2521-2530 (2000).
 Futaki et al., *J. Biol. Chem.* 276(8): 5836-40 (2001).
 Labrie et al., *Clin. Invest. Med.* 13(5): 275-8 (1990).
 Schafmeister et al. *J. Am. Chem. Soc.* 122: 5891-5892 (2000).
 Walenski et al., *Science* 305:1466-1470 (2004).
 Papo et al., *Cancer Res.* 64(16): 5779-86 (2004).
 Miller et al., *BiochemPharmacol.* 36(1): 169-76, (1987).
 Lee et al., *J. Pept. Res.* 63(2): 69-84 (2004).
 Monk et al., *BioDrugs* 19(4): 261-78 (2005).
 Defreest et al., *J. Pept. Res.* 63(5): 409-19 (2004).
 Toth et al., (*Developmental Cell* 1: 82-92 (2001)).
 Gough & Clothia, *Structure* 12: 917-925 (2004).
 De Rienzo et al., *Protein Science* 9: 1439-1454 (2000).
 Kuwabara, *Genes and Development* 17: 155-161 (2003).
 Murphy et al., *J. Mol. Biol.* 315: 859-871 (2002).
 Redinbo et al., *J. Bioenerg. Biomembr.* 26: 49-66 (1994).
 Walter, R.L. et al., *J. Mol. Biol.* 263: 730-51 (1996).
 Van Dreissche et al., *Protein Science* 8: 947-957 (1999).
 Bond et al., *J. Mol. Biol.* 306: 47-67 (2001).
 McManus et al. *J. Boil. Chem.* 267: 6531-6540 (1992).
 Koch et al. *J. Am. Chem. Soc.* 127: 158-166 (2005).
 Hart et al., *Protein Science* 5: 2175-2183 (1996).
 Guss et al., *J. Mol. Biol.* 262: 686-705 (1996).
 Cuzick, *Breast* 12: 405-411 (2003).
 Huang et al., *Am J. Epidemiol* 151: 703-714 (2000).
 Thiberville et al., *Cancer Res.* 55: 5133-5139 (1995).
 Mao et al., *J. Natl. Cancer Inst.* 89: 857-862 (1997).
 Tsao et al., *CA Clin. Cancer J. Clin.* 54: 150-180 (2004).
 Ulmer et al., *Science* 259: 1745-1749(1993).
 Kaufman, *Methods Enzymol.* 185: 487-511(1990).
 Carter, *Biochem J.* 237: 1-7 (1986).
 Zoller and Smith, *Methods Enzymol.* 154: 329-350 (1987).
 Wells et al. *Gene* 34: 315-323 (1985).
 Balin et al. *Skin & Aging*, Vol. Feb. 2001, pp. 37, 39, 40 and 43.
 Farver et al. *Israel Journal of Chemistry* 21: pp. 13-17 (1981).
 van Gastel et al. *J. Am. Chem. Soc.* 124(9) pp. 2035-2041 (2002).
 Office Action, dated Dec. 25, 2013, in Japanese Patent Application No. 2007-535767.
 Punj et al.: "Bacterial cupredoxin azurin as an inducer of apoptosis and regression in human breast cancer," *Oncogene*, vol. 23, pp. 2367-2378 (2004).
 Yamada et al.: "Bacterial redox protein azurin, tumor suppressor protein p53, and regression of cancer," *Proc. Natl. Acad. Sci.*, vol. 99, No. 22, pp. 14098-14103 (2002).

* cited by examiner

COMPARISON OF GLANDS WITH OR WITHOUT LESIONS IN AN ENLARGED PICTURE

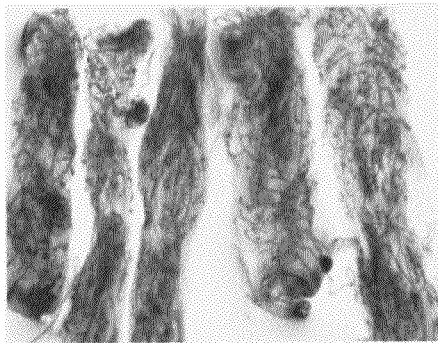
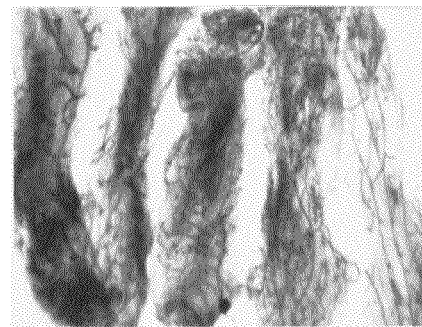


LESIONS



NO LESIONS

FIG. 1A

DMBA CONTROL 2 μ g/ml
$$+ \quad + \quad + \quad + \quad +$$

$$+ + + =$$

$$+ \quad + \quad + \quad + \quad \dots$$

INCIDENCE
12/15 80%

FIG. 1B

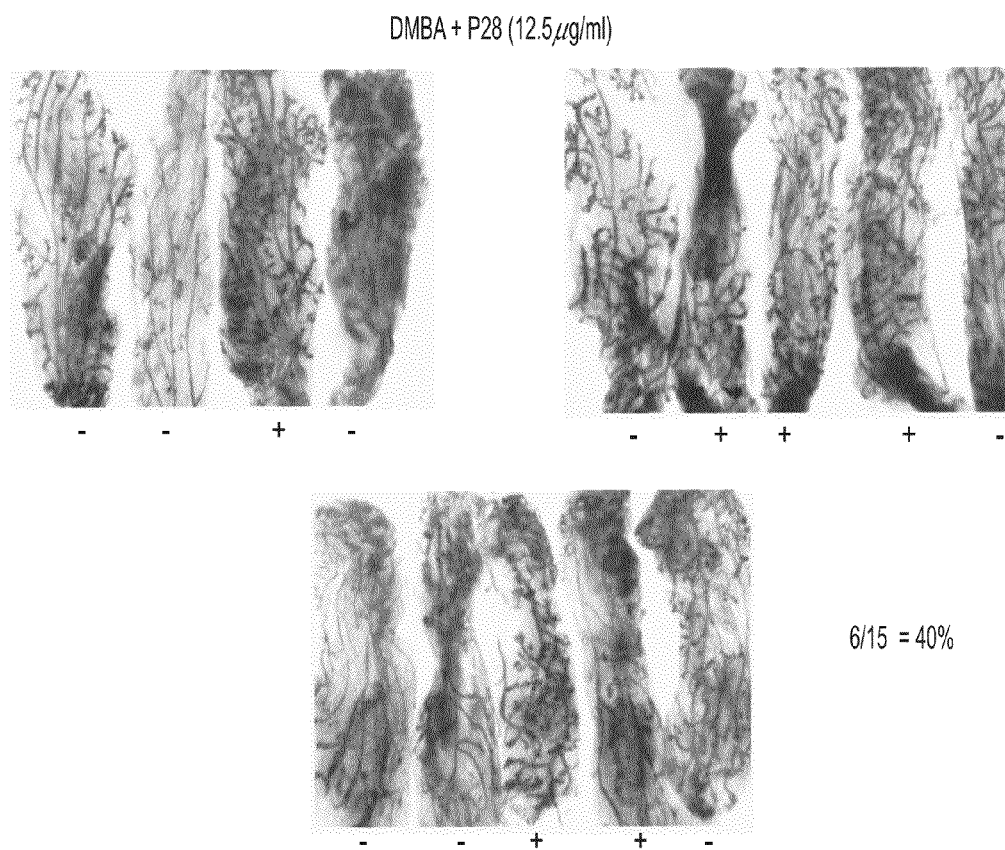


FIG. 1C

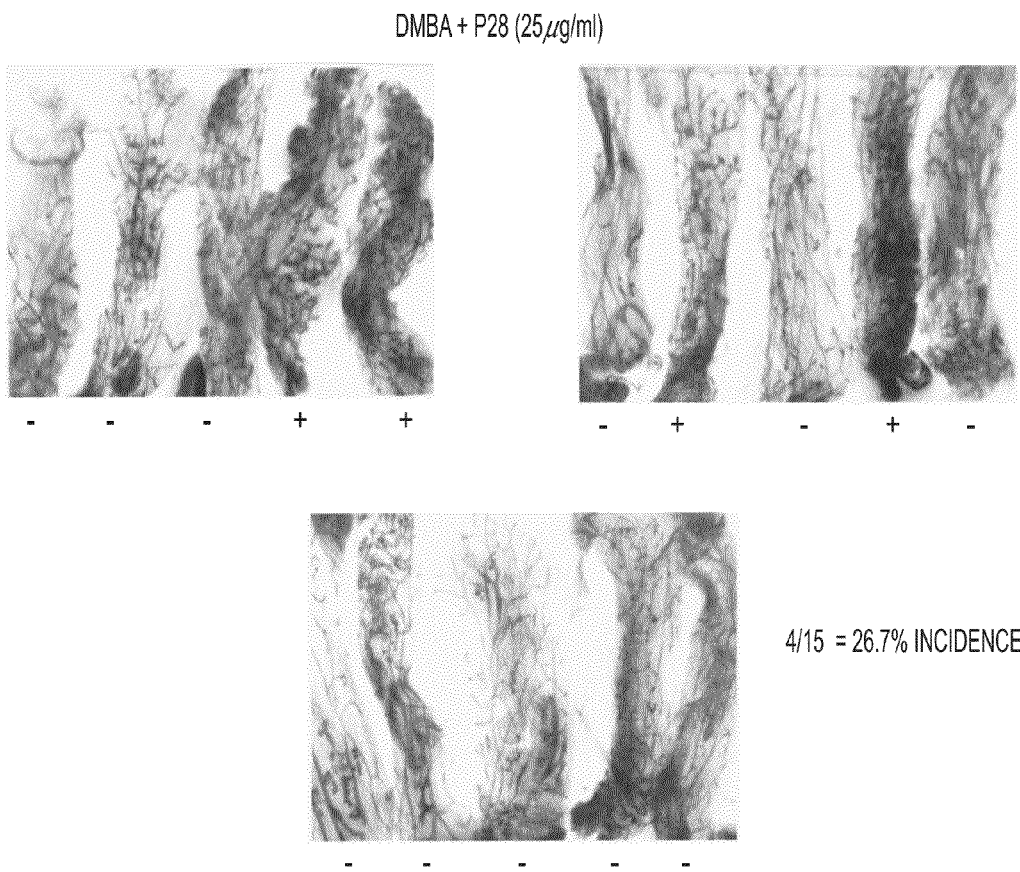


FIG. 1D

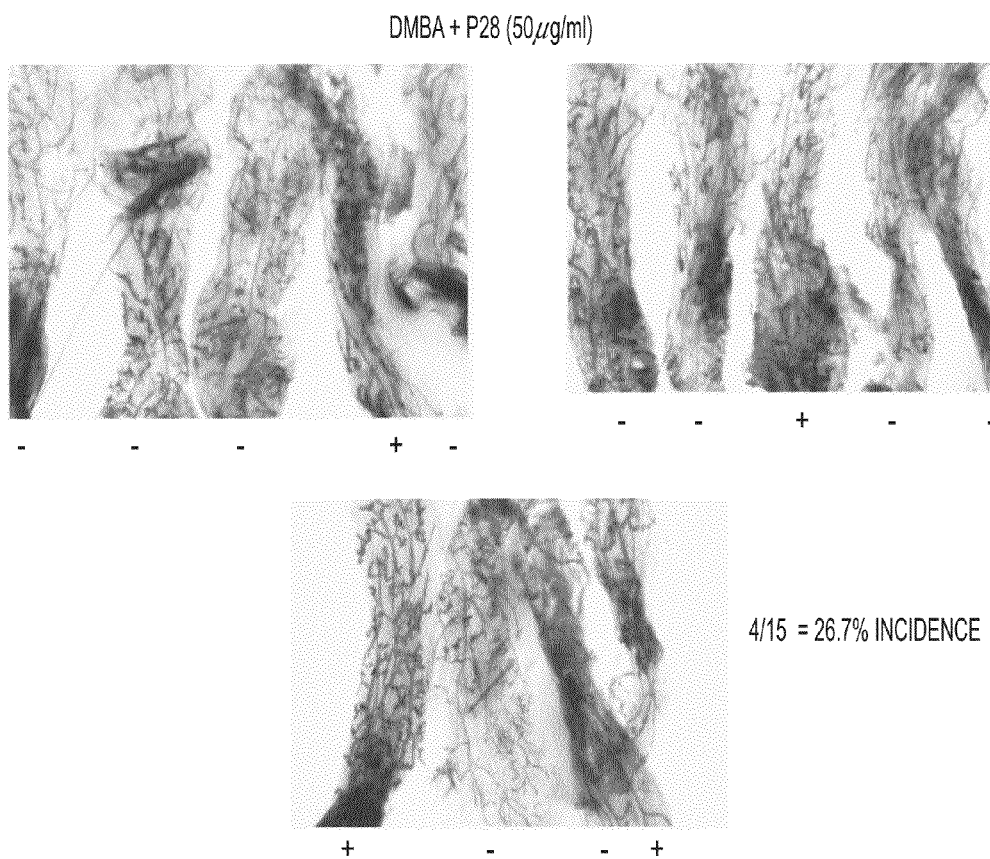


FIG. 1E

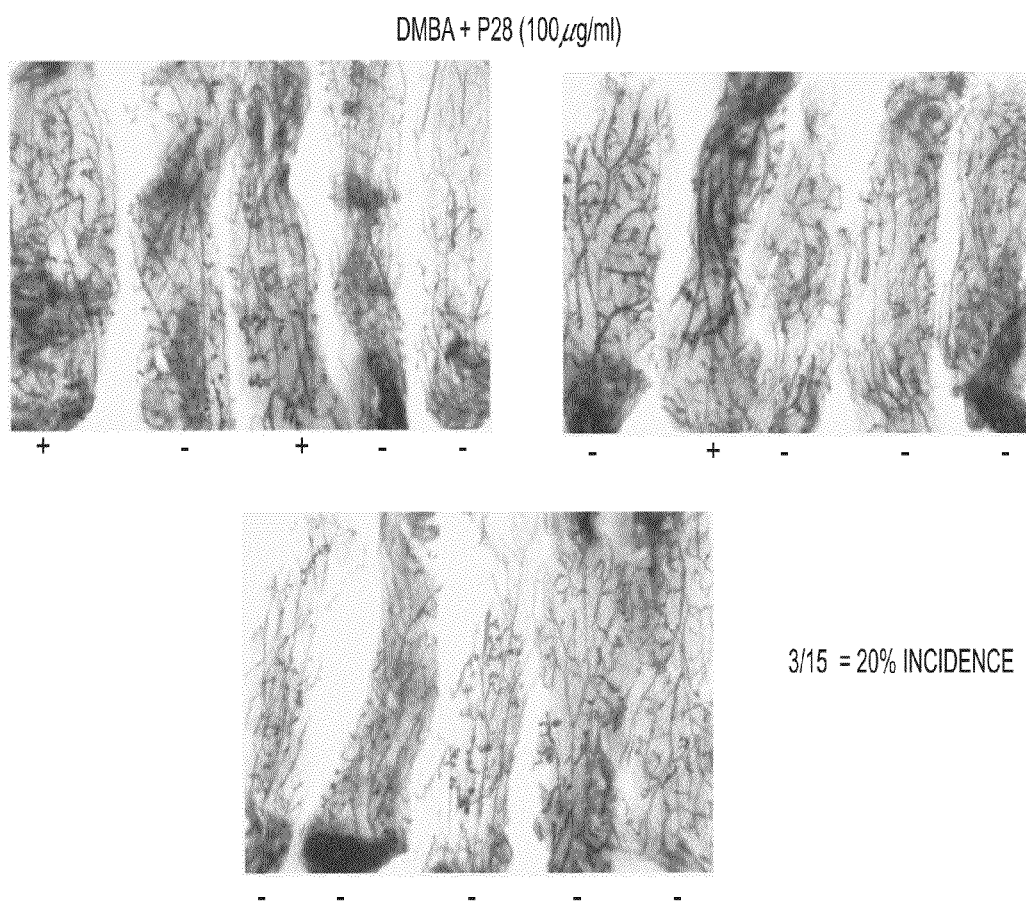


FIG. 1F

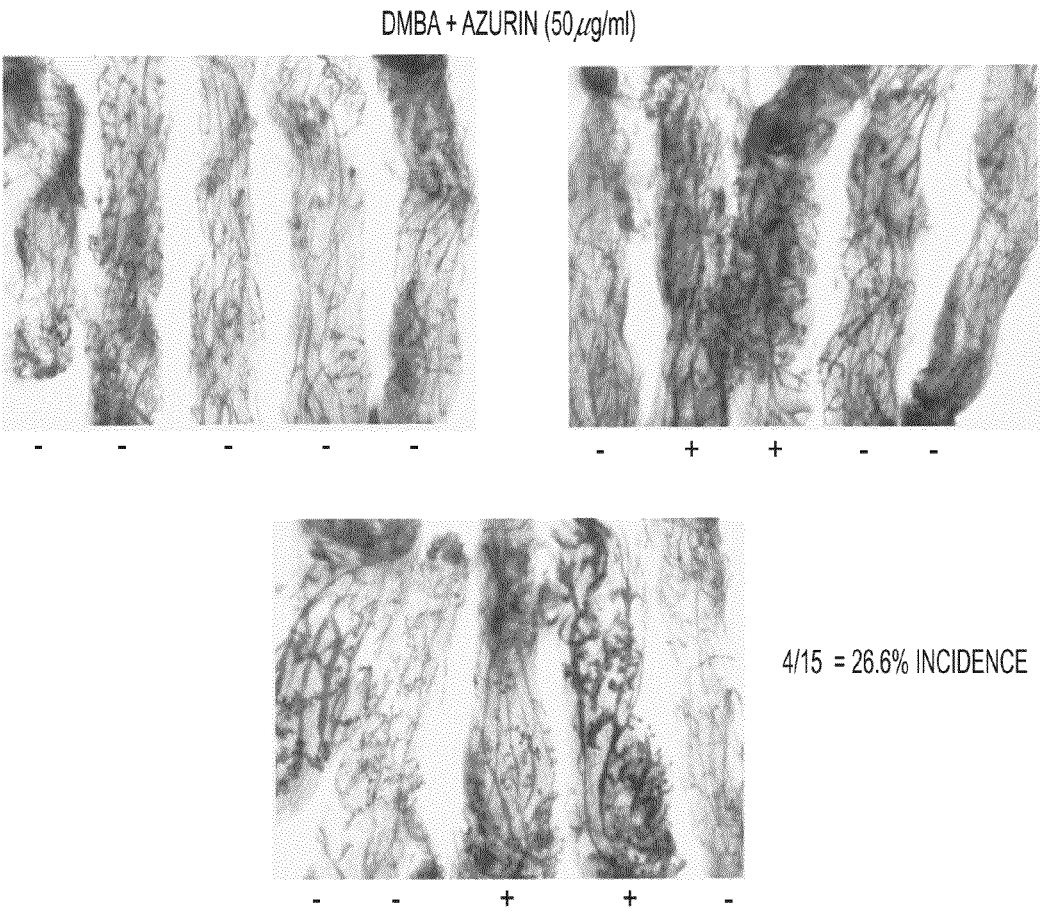


FIG. 1G

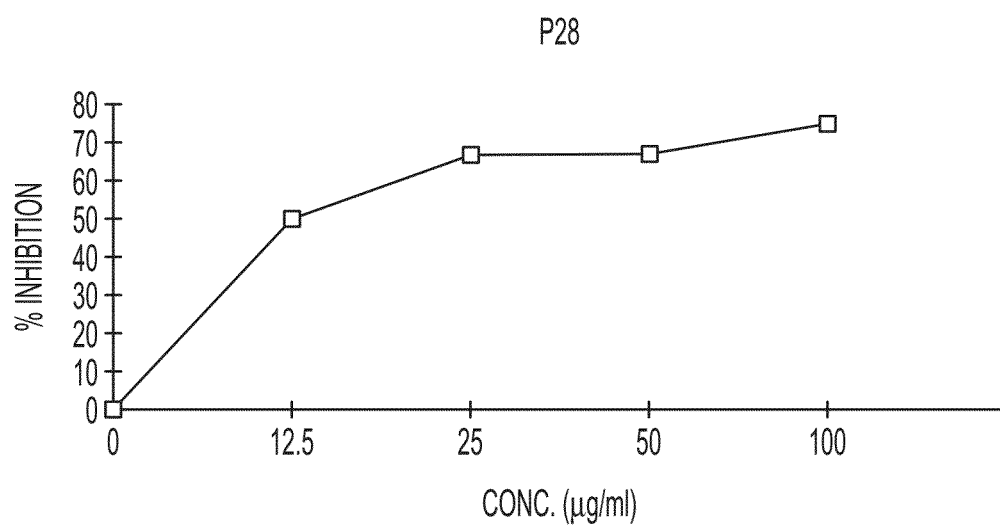
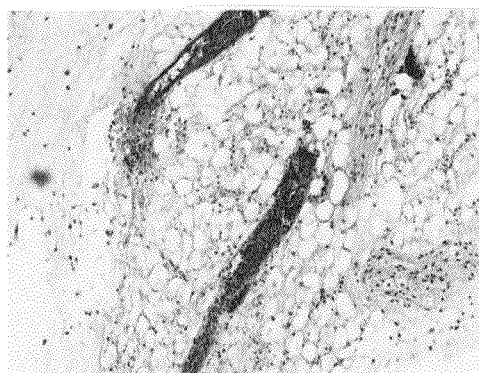
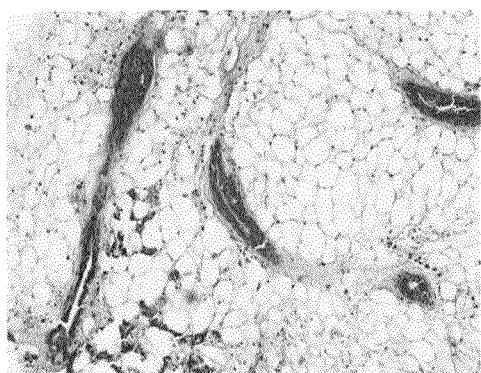


FIG. 2

EXAMPLES OF DMBA-INDUCED MAMMARY DUCTAL LESIONS



CONTROL DMBA ALONE

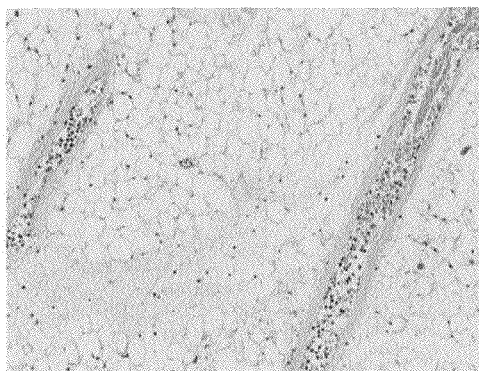
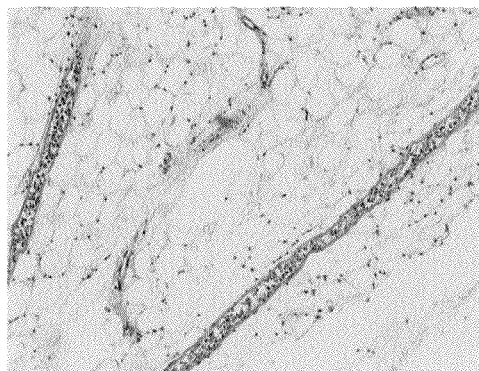
DMBA + AZURIN (50 μ g/ml)DMBA + P28 (50 μ g/ml)

FIG. 3

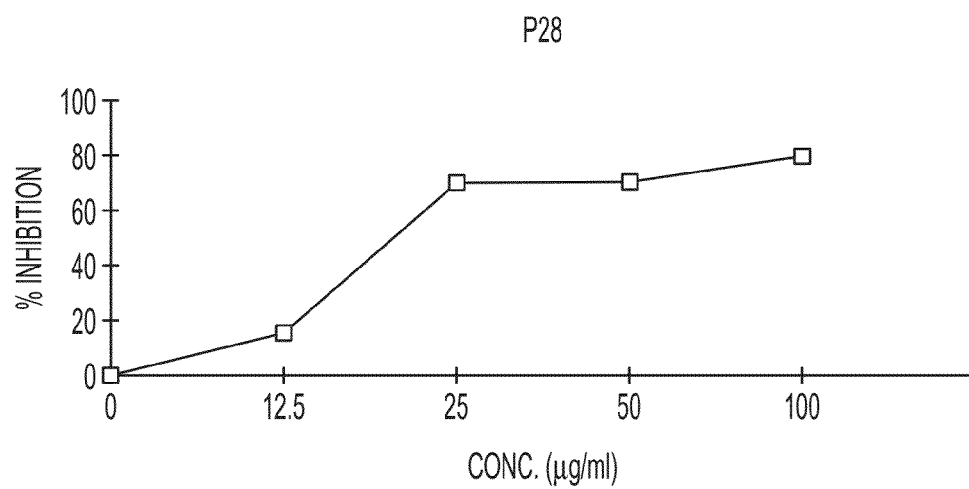


FIG. 4

COMPOSITIONS AND METHODS TO PREVENT CANCER WITH CUPREDOXINS

CROSS REFERENCE TO RELATED APPLICATIONS

This application claims priority under 35 U.S.C. §§119 and 120, to Provisional U.S. Application Ser. No. 60/844,358, filed Sep. 14, 2006; and is a continuation of U.S. patent application Ser. No. 12/617,841, filed Nov. 13, 2009, and is a continuation of U.S. patent application Ser. No. 11/854,654, filed Sep. 13, 2007, which issued as U.S. Pat. No. 7,618,939 on Nov. 17, 2009, which is a continuation-in-part of U.S. patent application Ser. No. 11/244,105, filed Oct. 6, 2005, which claims priority to U.S. Provisional Patent Application Ser. No. 60/616,782, filed Oct. 7, 2004, and U.S. Provisional Patent Application Ser. No. 60/680,500, filed May 13, 2005; and is a continuation-in-part of U.S. patent application Ser. No. 10/720,603, filed Nov. 24, 2003, which issued as U.S. Pat. No. 7,491,394 on Feb. 17, 2009, and which claims priority to U.S. Provisional Patent Application Ser. No. 60/414,550, filed Aug. 15, 2003, and which is a continuation-in-part of U.S. patent application Ser. No. 10/047,710, filed Jan. 15, 2002, which issued as U.S. Pat. No. 7,084,105 on Aug. 1, 2006, and which claims priority to U.S. Provisional Patent Application Ser. No. 60/269,133, filed Feb. 15, 2001. The entire content of these prior applications is fully incorporated herein by reference.

FIELD OF THE INVENTION

The present invention relates to compositions comprising variants, derivatives and structural equivalents of cupredoxins that inhibit the development of premalignant lesions in mammalian cells, tissues and animals. The invention also relates to the use of cupredoxins, and variants, derivatives and structurally equivalents of cupredoxins, as chemopreventive agents in mammals to inhibit the development of premalignant lesions, and ultimately cancer.

BACKGROUND

Cancer chemoprevention is the use of natural, synthetic or biologic chemical agents to reverse, suppress, or prevent carcinogenic progression to invasive cancer. Recent clinical trials in preventing cancer in high-risk populations suggest that chemopreventive therapy is a realistic treatment for high-risk patients. Chemopreventive therapy is based on the concepts of multifocal field carcinogenesis and multistep carcinogenesis. In field carcinogenesis, generalized carcinogen exposure throughout the tissue field results in diffuse epithelial injury in tissue and clonal proliferation of the mutated cells. These genetic mutations throughout the field increase the likelihood that one or more premalignant or malignant lesions may develop in the field. Multistep carcinogenesis in the stepwise accumulation of these genetic and phenotypic alterations. Arresting one or more steps in the multistep carcinogenesis may impede or prevent the development of cancer. See generally Tsao et al., *CA Cancer J Clin* 54:150-180 (2004).

Azurin, and other cupredoxins, are cytotoxic specifically towards cancer cells. Azurin induces apoptosis in J774 lung cancer cells. Yamada et al., *PNAS* 99(22):14098-14103 (2002). On entry into J774 lung cancer cells, azurin localizes in the cytosol and nuclear fractions, and forms a complex with tumor suppressor protein p53, thereby stabilizing it and enhancing its intracellular level. Id. The induction of azurin-mediated apoptosis is not limited to J774 cells. Azurin can

also enter cancer cells such as human melanoma UIISO-Mel-2 or human breast cancer MCF-7 cells. Yamada et al., *Infect Immun.* 70:7054-7062 (2002); Punj et al., *Oncogene*. 23:2367-2378 (2004). In both cases, azurin allowed the elevation of the intracellular p53 levels, leading to enhanced Bax formation and induction of apoptosis in such cells. Most interestingly, intraperitoneal injection of azurin in nude mice harboring xenografted Mel-2 or MCF-7 human cancers led to statistically significant regression of such cancers. Id.

The mouse mammary gland organ culture (MMOC) assay may be used to evaluate the inhibitory effects of potential chemopreventive agents on both hormone-induced structural differentiation of mammary glands and on the development of DMBA-induced preneoplastic hyperplastic alveolar nodule-like lesions in the gland. Mammary glands from young, virgin animals, when incubated for 6 days in the presence of insulin (I)+prolactin (P)+aldosterone (A), can differentiate into fully-grown glands. These glands morphologically resemble the glands obtained from pregnant mice. Aldosterone can be replaced by estrogen (E)+progesterone (Pg) Inclusion of hydrocortisone (H) to the medium stimulates the functional differentiation of the mammary glands. Mehta and Banerjee, *Acta Endocrinol.* 80:501 (1975); Mehta and Moon, *Breast Cancer: Treatment and Prognosis* 300, 300 (Basil A Stoll ed., Blackwell Press 1986). Thus, the hormone-induced structural and functional differentiation, observed in this culture system, mimics the responses to hormones observed during various physiological stages of the animal.

Mice exhibit a distinct preneoplastic stage prior to cancer formation in MMOC. Such preneoplastic lesions in C3H mice are induced by murine mammary tumor virus or in BALB/c mice by DMBA. Exposure of the glands to 2 µg/ml DMBA between days 3 and 4 of growth phases followed by regression of the glands for 2-3 weeks in the medium containing only insulin, results in the formation of mammary alveolar lesions (MAL). Hawthorne et al., *Pharmaceutical Biology* 40:70-74 (2002); Mehta et al., *Methods in Cell Science* 19:19-24 (1997). Furthermore, transplantation of epithelial cells, prepared from glands containing the DMBA-induced mammary lesions, into syngeneic host resulted in the development of mammary adenocarcinoma. Telang et al., *PNAS* 76:5886-5890 (1979). Pathologically, these tumors were similar to those observed in vivo when mice of the same strain are administered DMBA. Id.

DMBA-induced mammary lesion formation in MMOC can be inhibited by a variety of classes of chemopreventive agents such as retinoids. These agents include chemopreventive agents derived from the natural products such as brassinin and resveratrol, thiols, antioxidants, inhibitors of ornithine decarboxylase such as OFMO and deguelin, inhibitors of prostaglandin synthesis, Ca regulators, etc. Jang et al., *Science* 275:218-220 (1997); Mehta, *Eur. J. Cancer* 36:1275-1282 (2000); Metha et al., *J. Natl. Cancer Inst.* 89:212-219 (1997). These studies clearly demonstrate that this organ culture system offers a unique model to determine the effectiveness of compounds against mammary carcinogenesis. The results can be expected to closely correlate to the inhibition obtained by in vivo administration of such compounds.

The MMOC may also be induced to form mammary ductal lesions (MDL). The MDL can be induced if estrogen and progesterone instead of aldosterone and hydrocortisone are included in the medium. The alveolar structures in the presence of ovarian steroids are very small but the intraductal lesions are observed in histopathological sections. Mehta et al., *J. Natl. Cancer Inst.* 93:1103-1106 (2001). The antiestrogens, which selectively work on ovarian hormone dependent ER+ breast cancers such as tamoxifen, inhibited MDL for-

mation and not MAL. Thus, this modified culture model in addition to conventional MAL induction protocol now can be used to evaluate effects of chemopreventive agents on both MAL and MOL.

What is needed is a chemopreventive agent that inhibit the development of premalignant lesions. Such a chemopreventive agent should be able to either prevent the initial development of premalignant lesions, induce cell death in premalignant lesions that form, and/or prevent the development of premalignant lesions into malignant lesions. Such chemopreventive agents would have great utility in treating, in particular, patients who are at a high risk of developing cancer, due to either the presence of high-risk features, the presence of pre-malignant lesions, or the previous of cancer or premalignant lesions.

SUMMARY OF THE EMBODIMENTS

The present invention relates to compositions comprising peptides that may be variants, derivatives and structural equivalents of cupredoxins that inhibit the development of premalignant lesions in mammalian cells, tissues and animals. Specifically, these compositions may comprise azurin from *Pseudomonas aeruginosa*, and/or the 50-77 residue region of azurin (p28). The present invention further relates to compositions that may comprise cupredoxin(s), and/or variants, derivatives or structural equivalents of cupredoxins, that retain the ability to inhibit the development of premalignant lesions in mammalian cells, tissues or animals. These compositions may be isolated peptides or pharmaceutical compositions, among others. The compositions of the invention may be used in methods to prevent the development of cancer in mammalian patients.

One aspect of the invention are isolated peptides that may be a variant, derivative or structural equivalent of a cupredoxin; and may inhibit the development of premalignant lesions in mammalian tissue. The cupredoxin may be azurin, pseudoazurin, plastocyanin, rusticyanin, Laz, auracyanin, stellacyanin and cucumber basic protein, and specifically may be azurin. The cupredoxin may be from an organism such as *Pseudomonas aeruginosa*, *Alcaligenes faecalis*, *Ulva pertussis*, *Achromobacter xylosoxidans*, *Bordetella bronchiseptica*, *Methylobacter* sp., *Neisseria meningitidis*, *Neisseria gonorrhea*, *Pseudomonas fluorescens*, *Pseudomonas chlororaphis*, *Xylella fastidiosa* and *Vibrio parahaemolyticus*, and specifically may be *Pseudomonas aeruginosa*. In some embodiments, the peptide may be part of SEQ ID NOS: 1, 3-19, or has at least 80% amino acid sequence identity to SEQ ID NOS: 1, 3-19.

In some embodiments, the isolated peptide may be a truncation of a cupredoxin. The isolated peptide may be more than about 10 residues and not more than about 100 residues. The isolated peptide may comprise, or alternatively consist of, *Pseudomonas aeruginosa* azurin residues 50-77, *Pseudomonas aeruginosa* azurin residues 50-67, *Pseudomonas aeruginosa* azurin residues 36-88, or SEQ ID NOS: 20-24.

Another aspect of the invention is a pharmaceutical composition that may comprise at least one, or two, cupredoxins or isolated peptides of the invention in a pharmaceutically acceptable carrier. The pharmaceutical composition may be formulated for intravenous administration. In some embodiments, the cupredoxin in the pharmaceutical composition may be from an organism such as *Pseudomonas aeruginosa*, *Ulva pertussis*, *Alcaligenes faecalis*, *Achromobacter xylosoxidans*, *Bordetella bronchiseptica*, *Methylobacter* sp., *Neisseria meningitidis*, *Neisseria gonorrhea*, *Pseudomonas fluorescens*, *Pseudomonas chlororaphis*, *Xylella fastidiosa*

and *Vibrio parahaemolyticus*, and specifically may be from *Pseudomonas aeruginosa*. The cupredoxin may be SEQ ID NOS: 1, 3-19.

Another aspect of the invention is a method to treat a mammalian patient by administering to the patient a therapeutically effective amount of the pharmaceutical composition of the invention. The patient may be human, and may be at a higher risk to develop cancer than the general population. In some embodiments, the cancer may be melanoma, breast, pancreas, glioblastoma, astrocytoma, lung, colorectal, neck and head, bladder, prostate, skin, or cervical cancer. In some embodiments, the patient may have at least one high risk feature, premalignant lesions or have been cured of cancer or premalignant lesions.

The pharmaceutical composition may be administered by intravenous injection, intramuscular injection, subcutaneous injection, inhalation, topical administration, transdermal patch, suppository, vitreous injection or oral, and specifically may be administered by intravenous injection. The pharmaceutical composition may be co-administered with at least one other chemopreventive drug, and specifically at about the same time as another chemopreventive drug.

Another aspect of the invention is a kit comprising the pharmaceutical composition of the invention in a vial. The kit may be designed for intravenous administration.

Another aspect of the invention is a method to study the development of cancer comprising contacting mammalian cells with a cupredoxin or peptide of the invention and measuring the development of premalignant and malignant cells. In some embodiments, the cells may be human and/or mammary cells. In some embodiments, the cells are induced to develop premalignant lesions.

Another aspect of the invention is an expression vector, which encodes a peptide of the invention.

These and other aspects, advantages, and features of the invention will become apparent from the following figures and detailed description of the specific embodiments.

BRIEF DESCRIPTION OF THE SEQUENCES

SEQ ID NO: 1. Amino acid sequence of azurin from *Pseudomonas aeruginosa*.

SEQ ID NO: 2. Amino acid sequence of p28, *Pseudomonas aeruginosa* azurin residues 50-77.

SEQ ID NO: 3. Amino acid sequence of plastocyanin from *Phormidium laminosum*.

SEQ ID NO: 4. Amino acid sequence of rusticyanin from *Thiobacillus ferrooxidans*.

SEQ ID NO: 5. Amino acid sequence of pseudoazurin from *Achromobacter cycloclastes*.

SEQ ID NO: 6. Amino acid sequence of azurin from *Alcaligenes faecalis*.

SEQ ID NO: 7. Amino acid sequence of azurin from *Achromobacter xylosoxidans* ssp. *denitrificans* 1.

SEQ ID NO: 8. Amino acid sequence of azurin from *Bordetella bronchiseptica*.

SEQ ID NO: 9. Amino acid sequence of azurin from *Methylobacter* sp. J.

SEQ ID NO: 10. Amino acid sequence of azurin from *Neisseria meningitidis* Z2491.

SEQ ID NO: 11. Amino acid sequence of azurin from *Pseudomonas fluorescens*.

SEQ ID NO: 12. Amino acid sequence of azurin from *Pseudomonas chlororaphis*.

SEQ ID NO: 13. Amino acid sequence of azurin from *Xylella fastidiosa* 9a5c.

SEQ ID NO: 14. Amino acid sequence of stellacyanin from *Cucumis sativus*.

SEQ ID NO: 15. Amino acid sequence of auracyanin A from *Chloroflexus aurantiacus*.

SEQ ID NO: 16. Amino acid sequence of auracyanin B from *Chloroflexus aurantiacus*.

SEQ ID NO: 17. Amino acid sequence of cucumber basic protein from *Cucumis sativus*.

SEQ ID NO: 18. Amino acid sequence of Laz from *Neisseria gonorrhoeae* F62.

SEQ ID NO: 19. Amino acid sequence of the azurin from *Vibrio parahaemolyticus*.

SEQ ID NO: 20. Amino acid sequence of amino acids 57 to 89 of auracyanin B of *Chloroflexus aurantiacus*.

SEQ ID NO: 21. Amino acid sequence of amino acids 51-77 of *Pseudomonas syringae* azurin.

SEQ ID NO: 22. Amino acid sequence of amino acids 89-115 of *Neisseria meningitidis* Laz.

SEQ ID NO: 23. Amino acid sequence of amino acids 52-78 of *Vibrio parahaemolyticus* azurin.

SEQ ID NO: 24. Amino acid sequence of amino acids 51-77 of *Bordetella bronchiseptica* azurin.

SEQ ID NO: 25. Amino acid sequence of amino acids 50-67 of *Pseudomonas aeruginosa* azurin.

SEQ ID NO: 26. Amino acid sequence of amino acids 36-88 of *Pseudomonas aeruginosa* azurin.

BRIEF DESCRIPTION OF THE FIGURES

FIG. 1 depicts photographs of all of the glands evaluated for the efficacy of p28 and azurin. FIG. 1A shows a representative photograph of alveolar lesions in a DMBA-treated gland and its comparison with a gland that was treated with DMBA along with a chemopreventive agent. FIGS. 1B-1G show representative photographs of the effects of p28 on the development of alveolar lesions.

FIG. 2 depicts a graph showing the efficacy of p28 against DMBA-induced mammary alveolar lesions.

FIG. 3 depicts photographs of representative sections of ductal lesions and effect of p28.

FIG. 4 depicts a graph showing the efficacy of p28 against DMBA-induced ductal lesions

DETAILED DESCRIPTION OF THE INVENTION

Definitions

As used herein, the term "cell" includes either the singular or the plural of the term, unless specifically described as a "single cell."

As used herein, the terms "polypeptide," "peptide," and "protein" are used interchangeably to refer to a polymer of amino acid residues. The terms apply to amino acid polymers in which one or more amino acid residue is an artificial chemical analogue of a corresponding naturally occurring amino acid. The terms also apply to naturally occurring amino acid polymers. The terms "polypeptide," "peptide," and "protein" are also inclusive of modifications including, but not limited to, glycosylation, lipid attachment, sulfation, gamma-carboxylation of glutamic acid residues, hydroxylation and ADP-ribosylation. It will be appreciated that polypeptides are not always entirely linear. For instance, polypeptides may be branched as a result of ubiquitination and they may be circular (with or without branching), generally as a result of post-translation events, including natural processing event and events brought about by human manipulation which do not occur naturally. Circular, branched and

branched circular polypeptides may be synthesized by non-translation natural process and by entirely synthetic methods as well.

As used herein, the term "pharmacologic activity" means the effect of a drug or other chemical on a biological system. The effect of chemical may be beneficial (therapeutic) or harmful (toxic). The pure chemicals or mixtures may be of natural origin (plant, animal, or mineral) or may be synthetic compounds.

As used herein, the term "pre-malignant" means precancerous, or before abnormal cells divide without control.

As used herein, the term "lesion" means an area of abnormal tissue.

As used herein, the term "pathological condition" includes anatomic and physiological deviations from the normal that constitute an impairment of the normal state of the living animal or one of its parts, that interrupts or modifies the performance of the bodily functions, and is a response to various factors (as malnutrition, industrial hazards, or climate), to specific infective agents (as worms, parasitic protozoa, bacteria, or viruses), to inherent defects of the organism (as genetic anomalies), or to combinations of these factors.

As used herein, the term "condition" includes anatomic and physiological deviations from the normal that constitute an impairment of the normal state of the living animal or one of its parts, that interrupts or modifies the performance of the bodily functions.

As used herein, the term "suffering from" includes presently exhibiting the symptoms of a pathological condition, having a pathological condition even without observable symptoms, in recovery from a pathological condition, or recovered from a pathological condition.

As used herein, the term "chemoprevention" is the use of drugs, vitamins, or other agents to try to reduce the risk of, or delay the development or recurrence of, cancer.

As used herein, the term "treatment" includes preventing, lowering, stopping, or reversing the progression or severity of the condition or symptoms associated with a condition being treated. As such, the term "treatment" includes medical, therapeutic, and/or prophylactic administration, as appropriate. Treatment may also include preventing or lessening the development of a condition, such as cancer.

As used herein, the term "inhibit cell growth" means the slowing or ceasing of cell division and/or cell expansion. This term also includes the inhibition of cell development or increases in cell death.

A "therapeutically effective amount" is an amount effective to prevent, lower, stop or reverse the development of, or to partially or totally alleviate the existing symptoms of a particular condition for which the subject being treated. Determination of a therapeutically effective amount is well within the capability of those skilled in the art.

The term "substantially pure," as used herein, when used to modify a protein or other cellular product of the invention, refers to, for example, a protein isolated from the growth medium or cellular contents, in a form substantially free of, or unadulterated by, other proteins and/or other compounds. The term "substantially pure" refers to a factor in an amount of at least about 75%, by dry weight, of isolated fraction, or at least "75% substantially pure." More specifically, the term "substantially pure" refers to a compound of at least about 85%, by dry weight, of isolated fraction, or at least "85% substantially pure." Most specifically, the term "substantially pure" refers to a compound of at least about 95%, by dry weight, of isolated fraction, or at least "95% substantially pure." The term "substantially pure" may also be used to modify a synthetically-made protein or compound of the invention, where,

for example, the synthetic protein is isolated from the reagents and by-products of the synthesis reaction(s).

The term “pharmaceutical grade,” as used herein, when referring to a peptide or compound of the invention, is a peptide or compound that is isolated substantially or essentially from components which normally accompany the material as it is found in its natural state, including synthesis reagents and by-products, and substantially or essentially isolated from components that would impair its use as a pharmaceutical. For example, a “pharmaceutical grade” peptide may be isolated from any carcinogen. In some instances, “pharmaceutical grade” may be modified by the intended method of administration, such as “intravenous pharmaceutical grade,” in order to specify a peptide or compound that is substantially or essentially isolated from any substance that would render the composition unsuitable for intravenous administration to a patient. For example, an “intravenous pharmaceutical grade” peptide may be isolated from detergents, such as SDS, and anti-bacterial agents, such as azide.

The terms “isolated,” “purified” or “biologically pure” refer to material which is substantially or essentially free from components which normally accompany the material as it is found in its native state. Thus, isolated peptides in accordance with the invention preferably do not contain materials normally associated with the peptides in their in situ environment. An “isolated” region of a polypeptide refers to a region that does not include the whole sequence of the polypeptide from which the region was derived. An “isolated” nucleic acid, protein, or respective fragment thereof has been substantially removed from its in vivo environment so that it may be manipulated by the skilled artisan, such as but not limited to, nucleotide sequencing, restriction digestion, site-directed mutagenesis, and subcloning into expression vectors for a nucleic acid fragment as well as obtaining the protein or protein fragment in substantially pure quantities.

The term “variant” as used herein with respect to a peptide, refers to amino acid sequence variants which may have amino acids replaced, deleted, or inserted as compared to the wild-type polypeptide. Variants may be truncations of the wild-type peptide. A “deletion” is the removal of one or more amino acids from within the polypeptide, which a “truncation” is the removal of one or more amino acids from one or both ends of the polypeptide. Thus, a variant peptide may be made by manipulation of genes encoding the polypeptide. A variant may be made by altering the basic composition or characteristics of the polypeptide, but not at least some of its pharmacologic activities. For example, a “variant” of azurin can be a mutated azurin that retains its ability to inhibit the development of premalignant mammalian cells. In some cases, a variant peptide is synthesized with non-natural amino acids, such as ϵ -(3,5-dinitrobenzoyl)-Lys residues. Ghadiri & Fernholz, J. Am. Chem. Soc., 112:9633-9635 (1990). In some embodiments, the variant has not more than 20 amino acids replaced, deleted or inserted compared to wild-type peptide or part thereof. In some embodiments, the variant has not more than 15 amino acids replaced, deleted or inserted compared to wild-type peptide or part thereof. In some embodiments, the variant has not more than 10 amino acids replaced, deleted or inserted compared to wild-type peptide or part thereof. In some embodiments, the variant has not more than 6 amino acids replaced, deleted or inserted compared to wild-type peptide or part thereof. In some embodiments, the variant has not more than 5 amino acids replaced, deleted or inserted compared to wild-type peptide or part thereof. In some embodiments, the variant has not more than 3 amino acids replaced, deleted or inserted compared to wild-type peptide or part thereof.

The term “amino acid,” as used herein, means an amino acid moiety that comprises any naturally-occurring or non-naturally occurring or synthetic amino acid residue, i.e., any moiety comprising at least one carboxyl and at least one amino residue directly linked by one, two three or more carbon atoms, typically one (a) carbon atom.

The term “derivative” as used herein with respect to a peptide refers to a peptide that is derived from the subject peptide. A derivation includes chemical modifications of the peptide such that the peptide still retains some of its fundamental activities. For example, a “derivative” of azurin can, for example, be a chemically modified azurin that retains its ability to inhibit angiogenesis in mammalian cells. Chemical modifications of interest include, but are not limited to, amidation, acetylation, sulfation, polyethylene glycol (PEG) modification, phosphorylation or glycosylation of the peptide. In addition, a derivative peptide may be a fusion of a polypeptide or fragment thereof to a chemical compound, such as but not limited to, another peptide, drug molecule or other therapeutic or pharmaceutical agent or a detectable probe.

The term “percent (%) amino acid sequence identity” is defined as the percentage of amino acid residues in a polypeptide that are identical with amino acid residues in a candidate sequence when the two sequences are aligned. To determine % amino acid identity, sequences are aligned and if necessary, gaps are introduced to achieve the maximum % sequence identity; conservative substitutions are not considered as part of the sequence identity. Amino acid sequence alignment procedures to determine percent identity are well known to those of skill in the art. Often publicly available computer software such as BLAST, BLAST2, ALIGN2 or Megalign (DNASTAR) software is used to align peptide sequences. In a specific embodiment, Blastp (available from the National Center for Biotechnology Information, Bethesda Md.) is used using the default parameters of long complexity filter, expect 10, word size 3, existence 11 and extension 1.

When amino acid sequences are aligned, the % amino acid sequence identity of a given amino acid sequence A to, with, or against a given amino acid sequence B (which can alternatively be phrased as a given amino acid sequence A that has or comprises a certain % amino acid sequence identity to, with, or against a given amino acid sequence B) can be calculated as:

$$\% \text{ amino acid sequence identity} = X/Y * 100$$

where

X is the number of amino acid residues scored as identical matches by the sequence alignment program's or algorithm's alignment of A and B and

Y is the total number of amino acid residues in B.

If the length of amino acid sequence A is not equal to the length of amino acid sequence B, the % amino acid sequence identity of A to B will not equal the % amino acid sequence identity of B to A. When comparing longer sequences to shorter sequences, the shorter sequence will be the “B” sequence. For example, when comparing truncated peptides to the corresponding wild-type polypeptide, the truncated peptide will be the “B” sequence.

General

The present invention provides compositions comprising cupredoxin, and variants, derivatives and structural equivalents of cupredoxins, and methods to prevent the development of cancer in mammals. The invention also provides to variants, derivatives and structural equivalents of cupredoxin that retain the ability to prevent the development of cancer or the re-occurrence of cancer in mammals. Most particularly, the

invention provides compositions comprising *Pseudomonas aeruginosa* azurin, variants, derivatives and structural equivalents of azurin, and their use to treat patients, and particularly patients at a higher risk of developing cancer than the general population. Finally, the invention provides methods to study the development of cancer in mammalian cells, tissues and animals by contacting the cells with a cupredoxin, or variant, derivative or structural equivalent thereof, before or after inducing premalignant lesions, and observing the development of premalignant and/or malignant cells.

Previously, it was known that a redox protein elaborated by *Pseudomonas aeruginosa*, the cupredoxin azurin, selectively enters J774 lung cancer cells but not normal cells, and induces apoptosis. Zaborina et al., Microbiology 146:2521-2530 (2000). Azurin can also selectively enter and kill human melanoma UIISO-Mel-2 or human breast cancer MCF-7 cells. Yamada et al., PNAS 99:14098-14103 (2002); Punj et al., Oncogene 23:2367-2378 (2004). Azurin from *P. aeruginosa* preferentially enters J774 murine reticulum cell sarcoma cells, forms a complex with and stabilizes the tumor suppressor protein p53, enhances the intracellular concentration of p53, and induces apoptosis. Yamada et al., Infection and Immunity 70:7054-7062 (2002). Detailed studies of various domains of the azurin molecule showed that amino acids 50-77 (p28) (SEQ ID NO: 2) represented a protein transduction domain (PTD) critical for internalization and subsequent apoptotic activity. Yamada et al., Cell. Microbial. 7:1418-1431 (2005).

It is now known that azurin, and peptides derived from azurin, such as p28, have chemopreventive properties. It is now known that azurin, and p28, prevent to formation of premalignant preneoplastic lesions in mouse mammary gland organ culture. In a mouse mammary gland organ culture model, azurin at 50 µg/ml was found to inhibit the formation of alveolar lesions by 67%. Likewise, p28 at 25 µg/ml was found to inhibit the formation of alveolar lesions by 67%. See Example 1. Further, azurin at 50 µg/ml was found to inhibit the formation of ductal lesions by 79%, and p28 at 25 µg/ml inhibited the formation of ductal lesions by 71%. See Example 1. Confocal microscopy and FAC showed that azurin and p28 entered normal murine mammary epithelial cells (MM3MG) and mammary cancer cells (4T1). P28 also entered human umbilical vein endothelial cells (HUVEC) in a temperature, time and concentration dependent manner and inhibited capillary tube formation of HUVEC plated on Matrigel® in a dose dependent manner. It is therefore now known that azurin and variants of azurin may be used to inhibit the formation of premalignant preneoplastic lesions, and thus the development of cancer, and specifically breast cancer, in mammalian patients.

Due to the high degree of structural similarity between cupredoxins, it is likely that other cupredoxins will inhibit the formation of premalignant lesions in mammals as well as azurin. Such cupredoxins may be found in, for example, bacteria or plants. Several cupredoxins are known to have pharmacokinetic activities similar to those of azurin from *Pseudomonas aeruginosa*. For example, rusticyanin from *Thiobacillus ferrooxidans* can also enter macrophages and induce apoptosis. Yamada et al., Cell Cycle 3:1182-1187 (2004); Yamada et al., Cell. Micro. 7:1418-1431 (2005). Plastocyanin from *Phormidium laminosum* and pseudoazurin from *Achromobacter cycloclastes* also are cytotoxic towards macrophages. U.S. Pat. Pub. No. 20060040269, published Feb. 23, 2006. It is therefore contemplated that other cupredoxins may be used in the compositions and methods of the invention. Further, variants, derivatives, and structural equivalents of cupredoxins that retain the ability to inhibit the

formation of cancer in mammals may also be used in the compositions and methods of the invention. These variants and derivatives may include, but are not limited to, truncations of a cupredoxin, conservative substitutions of amino acids and proteins modifications such as PEGylation and all-hydrocarbon stabling of α -helices.

Compositions of the Invention

The invention provides for peptides that are variants, derivatives or structural equivalents of cupredoxin that inhibit the development of premalignant lesions in mammalian cells, tissues and animals. The invention further provides for peptides that are variants, derivatives or structural equivalents of cupredoxin that inhibit the development of cancer in mammalian cells, tissues and animals. In some embodiments, the peptide is isolated. In some embodiments, the peptide is substantially pure or pharmaceutical grade. In other embodiments, the peptide is in a composition that comprises, or consists essentially of, the peptide. In another specific embodiment, the peptide is non-antigenic and does not raise an immune response in a mammal, and more specifically a human. In some embodiments, the peptide is less than a full-length cupredoxin, and retains some of the pharmacologic activities of the cupredoxins. Specifically, in some embodiments, the peptide may retain the ability to inhibit the development of premalignant lesions in the mouse mammary gland organ culture.

The invention also provides compositions comprising at least one peptide that is a cupredoxin, or variant, derivative or structural equivalent of a cupredoxin, specifically in a pharmaceutical composition. In specific embodiments, the pharmaceutical composition is designed for a particular mode of administration, for example, but not limited to, oral, intraperitoneal, or intravenous. Such compositions may be hydrated in water, or may be dried (such as by lyophilization) for later hydration. Such compositions may be in solvents other than water, such as but not limited to, alcohol.

Because of the high structural homology between the cupredoxins, it is contemplated that cupredoxins will have the same chemopreventive properties as azurin and p28. In some embodiments, the cupredoxin is, but is not limited to, azurin, pseudoazurin, plastocyanin, rusticyanin, auracyanin, stellacyanin, cucumber basic protein or Laz. In particularly specific embodiments, the azurin is derived from *Pseudomonas aeruginosa*, *Alcaligenes faecalis*, *Achromobacter xylosoxidans* ssp. *denitrificans* I, *Bordetella bronchiseptica*, *Methylobacter* sp., *Neisseria meningitidis*, *Neisseria gonorrhoea*, *Pseudomonas fluorescens*, *Pseudomonas chlororaphis*, *Xylella fastidiosa*, *Ulva pertussis* or *Vibrio parahaemolyticus*. In a very specific embodiment, the azurin is from *Pseudomonas aeruginosa*. In other specific embodiments, the cupredoxin comprises an amino acid sequence that is SEQ ID NO: 1, 3-19.

The invention provides peptides that are amino acid sequence variants which have amino acids replaced, deleted, or inserted as compared to the wild-type cupredoxin. Variants of the invention may be truncations of the wild-type cupredoxin. In some embodiments, the peptide of the invention comprises a region of a cupredoxin that is less than the full length wild-type polypeptide. In some embodiments, the peptide of the invention comprises more than about 10 residues, more than about 15 residues or more than about 20 residues of a truncated cupredoxin. In some embodiments, the peptide comprises not more than about 100 residues, not more than about 50 residues, not more than about 40 residues, not more than about 30 residues or not more than about 20 residues of a truncated cupredoxin. In some embodiments, a cupredoxin has to the peptide, and more specifically SEQ ID NOS: 1,

3-19 as to the peptide of the invention, at least about 70% amino acid sequence identity, at least about 80% amino acid sequence identity, at least about 90% amino acid sequence identity, at least about 95% amino acid sequence identity or at least about 99% amino acid sequence identity.

In specific embodiments, the variant of cupredoxin comprises *P. aeruginosa* azurin residues 50-77 (p28, SEQ ID NO: 2), azurin residues 50-67, or azurin residues 36-88. In other embodiments, the variant of cupredoxin consists of *P. aeruginosa* azurin residues 50-77, azurin residues 50-67, or azurin residues 36-88. In other specific embodiments, the variant consists of the equivalent residues of a cupredoxin other than azurin. It is also contemplated that other cupredoxin variants can be designed that have a similar pharmacologic activity to azurin residues 50-77, or azurin residues 36-88. To do this, the subject cupredoxin amino acid sequence will be aligned to the *Pseudomonas aeruginosa* azurin sequence using BLAST, BLAST2, ALIGN2 or Megalign (DNASTAR), the relevant residues located on the *P. aeruginosa* azurin amino acid sequence, and the equivalent residues found on the subject cupredoxin sequence, and the equivalent peptide thus designed.

In one embodiment of the invention, the cupredoxin variant contains at least amino acids 57 to 89 of auracyanin B of *Chloroflexus aurantiacus* (SEQ ID NO: 20). In another embodiment of the invention, the cupredoxin variant contains at least amino acids 51-77 of *Pseudomonas syringae* azurin (SEQ ID NO: 21). In another embodiment of the invention, the cupredoxin variant contains at least amino acids 89-115 of *Neisseria meningitidis* Laz (SEQ ID NO: 22). In another embodiment of the invention, the cupredoxin variant contains at least amino acids 52-78 of *Vibrio parahaemolyticus* azurin (SEQ ID NO: 23). In another embodiment of the invention, the cupredoxin variant contains at least amino acids 51-77 of *Bordetella bronchiseptica* azurin (SEQ ID NO: 24).

The variants may also include peptides made with synthetic amino acids not naturally occurring. For example, non-naturally occurring amino acids may be integrated into the variant peptide to extend or optimize the half-life of the composition in the bloodstream. Such variants include, but are not limited to, D,L-peptides (diastereomer), (for example Futaki et al., J. Biol. Chem. 276(8):5836-40 (2001); Papo et al., Cancer Res. 64(16):5779-86 (2004); Miller et al, Biochem. Pharmacol. 36(1):169-76, (1987); peptides containing unusual amino acids (for example Lee et al., J. Pept. Res. 63(2):69-84 (2004)), olefin-containing non-natural amino acid followed by hydrocarbon stapling (for example Schafineister et al., J. Am. Chem. Soc. 122:5891-5892 (2000); Walenski et al., Science 305:1466-1470 (2004)), and peptides comprising ϵ -(3,5-dinitrobenzoyl)-Lys residues.

In other embodiments, the peptide of the invention is a derivative of a cupredoxin. The derivatives of cupredoxin are chemical modifications of the peptide such that the peptide still retains some of its fundamental activities. For example, a "derivative" of azurin can be a chemically modified azurin that retains its ability to inhibit the development of premalignant lesions in mammalian cells, tissues or animals. Chemical modifications of interest include, but are not limited to, hydrocarbon stapling, amidation, acetylation, sulfation, polyethylene glycol (PEG) modification, phosphorylation and glycosylation of the peptide. In addition, a derivative peptide maybe a fusion of a cupredoxin, or variant, derivative or structural equivalent thereof to a chemical compound, such as but not limited to, another peptide, drug molecule or other therapeutic or pharmaceutical agent or a detectable probe. Derivatives of interest include chemical modifications by which the half-life in the bloodstream of the peptides and

compositions of the invention can be extended or optimized, such as by several methods well known to those in the art, including but not limited to, circularized peptides (for example Monk et al., BioDrugs 19(4):261-78, (2005); DeFreest et al., J. Pept. Res. 63(5):409-19 (2004)), N- and C-terminal modifications (for example Labrie et al., Clin. Invest. Med. 13(5):275-8, (1990)), and olefin-containing non-natural amino acid followed by hydrocarbon stapling (for example Schafineister et al., J. Am. Chem. Soc. 122:5891-5892 (2000); Walenski et al., Science 305:1466-1470 (2004)).

In another embodiment, the peptide is a structural equivalent of a cupredoxin. Examples of studies that determine significant structural homology between cupredoxins and other proteins include Toth et al. (Developmental Cell 1:82-92 (2001)). Specifically, significant structural homology between a cupredoxin and the structural equivalent may be determined by using the VAST algorithm. Gibrat et al., Curr Opin Struct Biol 6:377-385 (1996); Madej et al., Proteins 23:356-3690 (1995). In specific embodiments, the VAST p value from a structural comparison of a cupredoxin to the structural equivalent may be less than about 10^{-3} , less than about 10^{-5} , or less than about 10^{-7} . In other embodiments, significant structural homology between a cupredoxin and the structural equivalent may be determined by using the DALI algorithm. Holm & Sander, J. Mol. Biol. 233:123-138 (1993). In specific embodiments, the DALI Z score for a pairwise structural comparison is at least about 3.5, at least about 7.0, or at least about 10.0.

It is contemplated that the peptides of the composition of invention may be more than one of a variant, derivative and/or structural equivalent of a cupredoxin. For example, the peptides may be a truncation of azurin that has been PEGylated, thus making it both a variant and a derivative. In one embodiment, the peptides of the invention are synthesized with α,α -disubstituted non-natural amino acids containing olefin-bearing tethers, followed by an all-hydrocarbon "staple" by ruthenium catalyzed olefin metathesis. Scharmeister et al., J. Am. Chem. Soc. 122:5891-5892 (2000); Walensky et al., Science 305:1466-1470 (2004). Additionally, peptides that are structural equivalents of azurin may be fused to other peptides, thus making a peptide that is both a structural equivalent and a derivative. These examples are merely to illustrate and not to limit the invention. Variants, derivatives or structural equivalents of cupredoxin may or may not bind copper.

In some embodiments, the cupredoxin, or variant, derivative or structural equivalent thereof has some of the pharmacologic activities of the *P. aeruginosa* azurin, and specifically p28. In a specific embodiment, the cupredoxins and variants, derivatives and structural equivalents of cupredoxins that may inhibit prevent the development of premalignant lesions in mammalian cells, tissues or animals, and specifically but not limited to, mammary gland cells. The invention also provides for the cupredoxins and variants, derivatives and structural equivalents of cupredoxins that may have the ability to inhibit the development of mammalian premalignant lesions, and specifically but not limited to, melanoma, breast, pancreas, glioblastoma, astrocytoma, lung, colorectal, neck and head, bladder, prostate, skin and cervical cancer cells. Inhibition of the development of cancer cells is any decrease, or lessening of the rate of increase, of the development of premalignant lesions that is statistically significant as compared to control treatments.

Because it is now known that cupredoxins can inhibit the development of premalignant lesions and ultimately cancer in mammalian cells, tissues or animals, and specifically breast cells, and more specifically, mouse mammary gland cells, it is

now possible to design variants and derivatives of cupredoxins that retain this chemopreventive activity. Such variants, derivatives and structural equivalents can be made by, for example, creating a "library" of various variants, derivatives and structural equivalents of cupredoxins and cupredoxin derived peptides and then testing each for chemopreventive activity, and specifically chemopreventive activity in the mouse mammary gland organ culture using one of many methods known in the art, such the exemplary method in Example 1. It is contemplated that the resulting variants, derivatives and structural equivalents of cupredoxins with chemopreventive activity may be used in the methods of the invention, in place of or in addition to azurin or p28.

In some specific embodiments, the variant, derivative or structural equivalent of cupredoxin may inhibit the development of 7,12-dimethylbenz (a) anthracene (DMBA) induced premalignant lesions in a mouse mammary gland organ culture (MMOC) to a degree that is statistically different from a non-treated control. A peptide can be tested for this activity by using the MMOC model system is described in Example 1, or as in Mehta et al. (J Natl Cancer Inst 93:1103-1106 (2001)) and Mehta et al. (Meth Cell Sci 19:19-24 (1997)). Other methods to determine whether cancer development is inhibited another are well known in the art and may be used as well.

In some specific embodiments, the variant, derivative or structural equivalent of cupredoxin inhibits the development of mammary alveolar lesions (MAL) in the a MMOC model to a degree that is statistically different from a non-treated

copper ion is solely bound by the protein matrix. A special distorted trigonal planar arrangement to two histidine and one cysteine ligands around the copper gives rise to very peculiar electronic properties of the metal site and an intense blue color. A number of cupredoxins have been crystallographically characterized at medium to high resolution.

The cupredoxins in general have a low sequence homology but high structural homology. Gough & Clothia, Structure 12:917-925 (2004); De Rienzo et al., Protein Science 9:1439-1454 (2000). For example, the amino acid sequence of azurin is 31% identical to that of auracyanin B, 16.3% to that of rusticyanin, 20.3% to that of plastocyanin, and 17.3% to that of pseudoazurin. See, Table 1. However, the structural similarity of these proteins is more pronounced. The VAST p value for the comparison of the structure of azurin to auracyanin B is $10^{-7.4}$, azurin to rusticyanin is 10^{-5} , azurin to plastocyanin is $10^{-5.6}$, and azurin to pseudoazurin is $10^{-4.1}$.

All of the cupredoxins possess an eight-stranded Greek key beta-barrel or beta-sandwich fold and have a highly conserved site architecture. De Rienzo et al., Protein Science 9:1439-1454 (2000). A prominent hydrophobic patch, due to the presence of many long chain aliphatic residues such as methionines and leucines, is present around the copper site in azurins, amicyanins, cyanobacterial plastocyanins, cucumber basic protein and to a lesser extent, pseudoazurin and eukaryotic plastocyanins Id. Hydrophobic patches are also found to a lesser extent in stellacyanin and rusticyanin copper sites, but have different features. Id.

TABLE 1

Sequence and structure alignment of azurin (1JZG) from <i>P. aeruginosa</i> to other proteins using VAST algorithm.						
PDB	Alignment length ¹	% aa identity	P-value ²	Score ³	⁽ⁱ⁾ RMSD ⁴	(ii) Description
1AOZ A 2	82	18.3	10e-7	12.2	1.9	Ascorbate oxidase
1QHQA	113	31	10e-7.4	12.1	1) 1.9	2) AuracyaninB
1V54 B 1	79	20.3	10e-6.0	11.2	2.1	Cytochrome c oxidase
1GY2 A	92	16.3	10e-5.0	11.1	3) 1.8	4) Rusticyanin
3MSP A	74	8.1	10e-6.7	10.9	2.5	Motile Major Sperm Protein ⁵
1IUZ	74	20.3	10e-5.6	10.3	5) 2.3	6) Plastocyanin
1KGY E	90	5.6	10e-4.6	10.1	7) 3.4	8) Ephrinb2
1PMY	75	17.3	10e-4.1	9.8	9) 2.3	10) Pseudoazurin

¹Aligned Length: The number of equivalent pairs of C-alpha atoms superimposed between the two structures, i.e. how many residues have been used to calculate the 3D superposition.

²P-VAL: The VAST p value is a measure of the significance of the comparison, expressed as a probability. For example, if the p value is 0.001, then the odds are 1000 to 1 against seeing a match of this quality by pure chance. The p value from VAST is adjusted for the effects of multiple comparisons using the assumption that there are 500 independent and unrelated types of domains in the MMDB database. The p value shown thus corresponds to the p value for the pairwise comparison of each domain pair, divided by 500.

³Score: The VAST structure-similarity score. This number is related to the number of secondary structure elements superimposed and the quality of that superposition. Higher VAST scores correlate with higher similarity.

⁴RMSD: The root mean square superposition residual in Angstroms. This number is calculated after optimal superposition of two structures, as the square root of the mean square distances between equivalent C-alpha atoms. Note that the RMSD value scales with the extent of the structural alignments and that this size must be taken into consideration when using RMSD as a descriptor of overall structural similarity.

⁵*C. elegans* major sperm protein proved to be an ephrin antagonist in oocyte maturation. Kuwabara, Genes and Development 17: 155-161 (2003).

control. In some specific embodiments, the variant, derivative or structural equivalent of cupredoxin inhibits the development of mammary ductal lesions (MDL) in the a MMOC model to a degree that is statistically different from a non-treated control. A peptide can be tested for these activities by using the MMOC model system induced to form premalignant lesions by DMBA, as described in Example 1. Evaluation of development of premalignant lesions in a MMOC model system may be determined by morphometric analysis, or histopathological analysis, as provided in Example 1.

Cupredoxins

These small blue copper proteins (cupredoxins) are electron transfer proteins (10-20 kDa) that participate in bacterial electron transfer chains or are of unknown function. The

Azurin

The azurins are copper containing proteins of 128 amino acid residues which belong to the family of cupredoxins involved in electron transfer in certain bacteria. The azurins include those from *P. aeruginosa* (PA) (SEQ ID NO: 1), *A. xylosoxidans*, and *A. denitrificans*. Murphy et al., J. Mol. Biol. 315:859-871 (2002). The amino acid sequence identity between the azurins varies between 60-90%, these proteins showed a strong structural homology. All azurins have a characteristic β -sandwich with Greek key motif and the single copper atom is always placed at the same region of the protein. In addition, azurins possess an essentially neutral hydrophobic patch surrounding the copper site. Id.

Plastocyanins

The plastocyanins are soluble proteins of cyanobacteria, algae and plants that contain one molecule of copper per molecule and are blue in their oxidized form. They occur in the chloroplast, where they function as electron carriers. Since the determination of the structure of poplar plastocyanin in 1978, the structure of algal (*Scenedesmus*, *Enteromorpha*, *Chlamydomonas*) and plant (French bean) plastocyanins has been determined either by crystallographic or NMR methods, and the poplar structure has been refined to 1.33 Å resolution. SEQ ID NO: 3 shows the amino acid sequence of plastocyanin from *Phormidium laminosum*, a thermophilic cyanobacterium. Another plastocyanin of interest is from *Ulva pertussis*.

Despite the sequence divergence among plastocyanins of algae and vascular plants (e.g., 62% sequence identity between the *Chlamydomonas* and poplar proteins), the three-dimensional structures are conserved (e.g., 0.76 Å rms deviation in the C alpha positions between the *Chlamydomonas* and *Poplar* proteins). Structural features include a distorted tetrahedral copper binding site at one end of an eight-stranded antiparallel beta-barrel, a pronounced negative patch, and a flat hydrophobic surface. The copper site is optimized for its electron transfer function, and the negative and hydrophobic patches are proposed to be involved in recognition of physiological reaction partners. Chemical modification, cross-linking, and site-directed mutagenesis experiments have confirmed the importance of the negative and hydrophobic patches in binding interactions with cytochrome f, and validated the model of two functionally significant electron transfer paths involving plastocyanin. One putative electron transfer path is relatively short (approximately 4 Å) and involves the solvent-exposed copper ligand His-87 in the hydrophobic patch, while the other is more lengthy (approximately 12-15 Å) and involves the nearly conserved residue Tyr-83 in the negative patch. Redinbo et al., J. Bioenerg. Biomembr. 26:49-66 (1994).

Rusticyanins

Rusticyanins are blue-copper containing single-chain polypeptides obtained from a *Thiobacillus* (now called *Acidithiobacillus*). The X-ray crystal structure of the oxidized form of the extremely stable and highly oxidizing cupredoxin rusticyanin from *Thiobacillus ferrooxidans* (SEQ ID NO: 4) has been determined by multiwavelength anomalous diffraction and refined to 1.9 Å resolution. The rusticyanins are composed of a core beta-sandwich fold composed of a six- and a seven-stranded b-sheet. Like other cupredoxins, the copper ion is coordinated by a cluster of four conserved residues (His 85, Cys138, His143, Met148) arranged in a distorted tetrahedron. Walter, R. L. et al., J. Mol. Biol. 263: 730-51 (1996).

Pseudoazurins

The pseudoazurins are a family of blue-copper containing single-chain polypeptide. The amino acid sequence of pseudoazurin obtained from *Achromobacter cycloclastes* is shown in SEQ ID NO: 5. The X-ray structure analysis of pseudoazurin shows that it has a similar structure to the azurins although there is low sequence homology between these proteins. Two main differences exist between the overall structure of the pseudoazurins and azurins. There is a carboxy terminus extension in the pseudoazurins, relative to the azurins, consisting of two alpha-helices. In the mid-peptide region azurins contain an extended loop, shortened in the pseudoazurins, which forms a flap containing a short α -helix. The only major differences at the copper atom site are the confor-

mation of the MET side-chain and the Met-S copper bond length, which is significantly shorter in pseudoazurin than in azurin.

Phytocyanins

The proteins identifiable as phytocyanins include, but are not limited to, cucumber basic protein, stellacyanin, mavecyanin, umecyanin, a cucumber peeling cupredoxin, a putative blue copper protein in pea pods, and a blue copper protein from *Arabidopsis thaliana*. In all except cucumber basic protein and the pea-pod protein, the axial methionine ligand normally found at blue copper sites is replaced by glutamine. Auracyanin

Three small blue copper proteins designated auracyanin A, auracyanin B-1, and auracyanin B-2 have been isolated from the thermophilic green gliding photosynthetic bacterium *Chloroflexus aurantiacus*. The two B forms are glycoproteins and have almost identical properties to each other, but are distinct from the A form. The sodium dodecyl sulfate-polyacrylamide gel electrophoresis demonstrates apparent monomer molecular masses as 14 (A), 18 (B-2), and 22 (B-1) kDa.

The amino acid sequence of auracyanin A has been determined and showed auracyanin A to be a polypeptide of 139 residues. Van Dreissche et al., Protein Science 8:947-957 (1999). His58, Cys123, His128, and Met132 are spaced in a way to be expected if they are the evolutionary conserved metal ligands as in the known small copper proteins plastocyanin and azurin. Secondary structure prediction also indicates that auracyanin has a general beta-barrel structure similar to that of azurin from *Pseudomonas aeruginosa* and plastocyanin from poplar leaves. However, auracyanin appears to have sequence characteristics of both small copper protein sequence classes. The overall similarity with a consensus sequence of azurin is roughly the same as that with a consensus sequence of plastocyanin, namely 30.5%. The N-terminal sequence region 1-18 of auracyanin is remarkably rich in glycine and hydroxy amino acids. Id. See exemplary amino acid sequence SEQ ID NO: 15 for chain A of auracyanin from *Chloroflexus aurantiacus* (NCBI Protein Data Bank Accession No. AAM12874).

The auracyanin B molecule has a standard cupredoxin fold. The crystal structure of auracyanin B from *Chloroflexus aurantiacus* has been studied. Bond et al., J. Mol. Biol. 306: 47-67 (2001). With the exception of an additional N-terminal strand, the molecule is very similar to that of the bacterial cupredoxin, azurin. As in other cupredoxins, one of the Cu ligands lies on strand 4 of the polypeptide, and the other three lie along a large loop between strands 7 and 8. The Cu site geometry is discussed with reference to the amino acid spacing between the latter three ligands. The crystallographically characterized Cu-binding domain of auracyanin B is probably tethered to the periplasmic side of the cytoplasmic membrane by an N-terminal tail that exhibits significant sequence identity with known tethers in several other membrane-associated electron-transfer proteins. The amino acid sequences of the B forms are presented in McManus et al. J. Biol. Chem. 267:6531-6540 (1992). See exemplary amino acid sequence SEQ ID NO: 16 for chain B of auracyanin from *Chloroflexus aurantiacus* (NCBI Protein Data Bank Accession No. 1QHQA).

Stellacyanin

Stellacyanins are a subclass of phytocyanins, a ubiquitous family of plant cupredoxins. An exemplary sequence of a stellacyanin is included herein as SEQ ID NO: 14. The crystal structure of umecyanin, a stellacyanin from horseradish root (Koch et al., J. Am. Chem. Soc. 127:158-166 (2005)) and cucumber stellacyanin (Hart et al., Protein Science 5:2175-2183 (1996)) is also known. The protein has an overall fold

similar to the other phytochemicals. The ephrin B2 protein ectodomain tertiary structure bears a significant similarity to stellacyanin. Toth et al., *Developmental Cell* 1:83-92 (2001). An exemplary amino acid sequence of a stellacyanin is found in the National Center for Biotechnology Information Protein Data Bank as Accession No. 1JER, SEQ ID NO: 14. Cucumber Basic Protein

An exemplary amino acid sequence from a cucumber basic protein is included herein as SEQ ID NO: 17. The crystal structure of the cucumber basic protein (CBP), a type 1 blue copper protein, has been refined at 1.8 Å resolution. The molecule resembles other blue copper proteins in having a Greek key beta-barrel structure, except that the barrel is open on one side and is better described as a "beta-sandwich" or "beta-taco". Guss et al., *J. Mol. Biol.* 262:686-705 (1996). The ephrinB2 protein ectodomain tertiary structure bears a high similarity (rms deviation 1.5 Å for the 50α carbons) to the cucumber basic protein. Toth et al., *Developmental Cell* 1:83-92 (2001).

The Cu atom has the normal blue copper NNSS' co-ordination with bond lengths Cu—N(His39)=1.93 Å, Cu—S(Cys79)=2.16 Å, Cu—N(His84)=1.95 Å, Cu—S(Met89)=2.61 Å. A disulphide link, (Cys52)—S—S—(Cys85), appears to play an important role in stabilizing the molecular structure. The polypeptide fold is typical of a sub-family of blue copper proteins (phytochemicals) as well as a non-metalloprotein, ragweed allergen Ra3, with which CBP has a high degree of sequence identity. The proteins currently identifiable as phytochemicals are CBP, stellacyanin, macyanin, umecyanin, a cucumber peeling cupredoxin, a putative blue copper protein in pea pods, and a blue copper protein from *Arabidopsis thaliana*. In all except CBP and the pea-pod protein, the axial methionine ligand normally found at blue copper sites is replaced by glutamine. An exemplary sequence for cucumber basic protein is found in NCBI Protein Data Bank Accession No. 2CBP, SEQ ID NO: 17.

Methods of Use

The invention provides methods to prevent de novo malignancies in otherwise healthy patients comprising administering to the patient at least one peptide that is a cupredoxin, or variant, derivative or structural equivalent thereof, as described above. Chemopreventive therapies are based on the hypothesis that the interruption of processes involved in cancerogenesis will prevent the development of cancer. The cupredoxin *Pseudomonas aeruginosa* azurin and the truncated azurin peptide p28 are now known to inhibit the development of premalignant lesions, either by inhibiting the initial formation of premalignant lesions, or killing or inhibiting the growth of premalignant lesions that are present. It therefore contemplated that a cupredoxin, or variant, derivative or structural equivalent thereof, as described above, with the ability to inhibit the development of premalignant lesions, may be used in chemopreventive therapies in otherwise healthy patients. Such otherwise healthy patients are, in some embodiments, patients at a higher risk to develop cancer than those in the general population. Cancers that may be prevented by treatment with the compositions of the invention include, but are not limited to, melanoma, breast, pancreas, glioblastoma, astrocytoma, lung, colorectal, neck and head, bladder, prostate, skin, and cervical cancer. In some embodiments, the patient may be human. In other embodiments, the patient is not human.

The invention further includes methods to study the development of cancer comprising contacting mammalian cells before or after induction with a carcinogen with a composition comprising cupredoxin, or a variant, derivative or structural equivalent thereof and observing the development of the

cells. In some embodiments, the cells are mouse mammary gland cells, while in others they are other cells that may become malignant in mammals.

Patients at a higher at risk to develop cancer than the general population may be patients with high risk features, patients with premalignant lesions, and patients that have been cured of their initial cancer or definitively treated for their premalignant lesions. See generally Tsao et al., *CA Cancer J Clin* 54:150-180 (2004). High risk features may be behavioral, genetic, environmental or physiological factors of the patient. Behavioral factors that predispose a patient to various forms of cancer include, but are not limited to, smoking, diet, alcohol consumption, hormone replacement therapy, higher body mass index, nulliparity, beta1 nut use, frequent mouthwash use, exposure to human papillomavirus, childhood and chronic sun exposure, early age of first intercourse, multiple sexual partners, and oral contraceptive use. Genetic factors that predispose a patient to various forms of cancer include, but are not limited to, a family history of cancer, gene carrier status of BRCA/and BRCA2, prior history of breast neoplasia, familial adenomatous polyposis (FAP), hereditary nonpolyposis colorectal cancer (HNPCC), red or blond hair and fair-skinned phenotype, xeroderma pigmentosum, and ethnicity. Environmental features that predispose a patient to various forms of cancer include, but are not limited to, exposure to radon, polycyclic aromatic hydrocarbons, nickel, chromate, arsenic, asbestos, chloromethyl ethers, benzo[a]pyrene, radiation, and aromatic amines from rubber or paint occupational exposure. Other miscellaneous factors that predispose a patient to various forms of cancer include, but are not limited to, chronic obstructive pulmonary disease with airflow obstruction, chronic bladder infections, schistosomiasis, older age, and immunocompromised status.

Additionally, patients at a higher risk of developing cancer may be determined by the use of various risk models that have been developed for certain kinds of cancer. For example, patients predisposed to breast cancer may be determined using the Gail risk model, or the Claus model, among others. See Gail et al., *J Natl Cancer Inst* 81:1879-1886 (1989); Cuzick, *Breast* 12:405-411 (2003); Huang et al., *Am J Epidemiol* 151:703-714 (2000).

Patients with premalignant lesions are at a higher risk to develop cancer than the general population. The presence of premalignant lesions in or on a patient may be determined by many methods that are well known to those in the art. Intermediate markers or biomarkers that originate from premalignant lesions may be measured in a patient to determine if the patient harbors premalignant lesions. Chromosomal abnormalities occur in tumor cells and the adjacent histologically normal tissues in the majority of cancer patients. Progression in chromosomal abnormalities parallels the phenotypic progression from premalignant lesion to invasive cancer. Thiberville et al., *Cancer Res.* 55:5133-5139 (1995). Therefore, chromosomal abnormalities associated with cancer may be used as intermediate markers to detect premalignant lesions in a patient. Common chromosomal abnormalities associated with cancer include, but are not limited to, allelic deletions or loss of heterozygosity (LOH) in tumor suppressor genes such as 3p (FHIT and others), 9p (9p21 for p16^{INK4}, p15^{INK4B}, and p19^{ARF}), 17p (17p13 for p53 gene and others) and 13q (13q14 for retinoblastoma gene Rb and others). Deletions in 3p and 9p are associated with smoking and the early stages of lung cancer. Mao et al., *J. Natl. Cancer Inst.* 89:857-862 (1997). Deletions affecting 3p, 5q, 8p, 17p and 18q are common change in epithelial cancers. See generally Tsao et al., *CA Clin. Cancer J. Clin.* 54:153 (2004). Other chromosomal mutations associated with cancer include those which acti-

vate oncogenes. Oncogenes whose presence may be used as intermediate markers include, but are not limited to, Ras, c-myc, epidermal growth factor, erb-B2 and cyclins E, D1 and B1. See generally id. at 154.

Other intermediate markers may be the products of genes up-regulated in premalignant cells and cancer cells. Genes that may be up-regulated in premalignant cells include, but are not limited to, cyclooxygenases COX-1 and COX-2, telomerase. Other biomarkers of cancer cells, and some premalignant cells, include, but are not limited to, p53, epidermal growth factor receptor (GFR), proliferating cell nuclear antigen (PCNA), RAS, COX-2, Ki-67, DNA aneuploidy, DNA polymerase- α , ER, Her2neu, E-cadherin, RAR β , hTERT, p16^{INK4a}, FHIT (3p14), Bcl-2, VEGF-R, HPV infection, LOH 9p21, LOH 17p, p-AKT, hnRNP A2/B1, RAF, Myc, c-KIT, cyclin D1, E and B1, IGF1, bcl-2, p16, LOH 3p21.3, LOH 3p2.5, LOH 9p21, LOH 17p13, LOH 13q, LOH 8p, hMSH2, APC, DCC, DPC4, Jv18, BAX, PSA, GSTP1, NF-kB, AP1, D3S2, HPV infection, LOH 3p14, LOH 4q, LOH 0.5p, bladder tumor antigen (BTA), BTK TRAK (Alidex, Inc., Redmond Wash.), urinary tract matrix protein 22, fibrin degradation product, autodrine motility factor receptor, BCLA-4, cytokeratin 20, hyaluronic acid, CYFRA 21-1, BCA, beta-human chorionic gonadotropin, and tissue polypeptide antigen (TPA). See generally id. at 155-157.

Patients that have been cured of their initial cancers or have been definitively treated for their premalignant lesions are also at a higher risk to develop cancer than the general population. A second primary tumor refers to a new primary cancer in a person with a history of cancer. Second primary tumors are the leading cause of mortality in head and neck cancer. Id. at 150. A second primary tumor is distinct from a metastasis in that the former originates de novo while the later originates from an existing tumor. Patients that have been cured of cancer or premalignant lesions of the breast, head and neck, lung, and skin are at a particularly high risk to develop second primary tumors.

The compositions comprising a cupredoxin or variant, derivative or structural equivalent thereof can be administered to the patient by many routes and in many regimens that will be well known to those in the art. In specific embodiments, the cupredoxin, or variant, derivative or structural equivalent thereof is administered intravenously, intramuscularly, subcutaneously, topically, orally, or by inhalation. The compositions may be administered to the patient by any means that delivers the peptides to the site in the patient that is at risk of developing cancer. In specific embodiments, the cupredoxin or variant, derivative or structural equivalent thereof is administered intravenously.

In one embodiment, the methods may comprise co-administering to a patient one unit dose of a composition comprising a cupredoxin or a variant, derivative or structural equivalent of cupredoxin and one unit dose of a composition comprising another chemopreventive drug, in either order, administered at about the same time, or within about a given time following the administration of the other, for example, about one minute to about 60 minutes following the administration of the other drug, or about 1 hour to about 12 hours following the administration of the other drug. Chemopreventive drugs of interest include, but are not limited to, tamoxifen, aromatase inhibitors such as letrozole and anastrozole (Arimidex®), retinoids such as N-[4-hydroxyphenyl]retinamide (4-HPR, fenretinide), nonsteroidal antiinflammatory agents (NSAIDs) such as aspirin and sulindac, celecoxib (COX-2 inhibitor), defluoromethylornithing (DFMO), ursodeoxycholic acid, 3-hydroxy-3-methylglutaryl coenzyme A reductase inhibitors, EKI-785 (EGFR inhibitor), bevacizumab (antibody to

VEGF-receptor), cetuximab (antibody to EGFR), retinol such as vitamin A, beta-carotene, 13-cis retinoic acid, isotretinoin and retinyl palmitate, α -tocopherol, interferon, oncolytic adenovirus d11520 (ONYX-015), gefitinib, etretinate, finasteride, indole-3-carbinol, resveratrol, chlorogenic acid, raloxifene, and olipraz.

Pharmaceutical Compositions Comprising Cupredoxin, or Variant, Derivative or Structural Equivalent Thereof

Pharmaceutical compositions comprising cupredoxin or variant, derivative or structural equivalents thereof, can be manufactured in any conventional manner, e.g., by conventional mixing, dissolving, granulating, dragee-making, emulsifying, encapsulating, entrapping, or lyophilizing processes. The substantially pure or pharmaceutical grade cupredoxin or variants, derivatives and structural equivalents thereof can be readily combined with a pharmaceutically acceptable carrier well-known in the art. Such carriers enable the preparation to be formulated as a tablet, pill, dragee, capsule, liquid, gel, syrup, slurry, suspension, and the like. Suitable carriers or excipients can also include, for example, fillers and cellulose preparations. Other excipients can include, for example, flavoring agents, coloring agents, detackifiers, thickeners, and other acceptable additives, adjuvants, or binders. In some embodiments, the pharmaceutical preparation is substantially free of preservatives. In other embodiments, the pharmaceutical preparation may contain at least one preservative. General methodology on pharmaceutical dosage forms is found in Ansel et al., *Pharmaceutical Dosage Forms and Drug Delivery Systems* (Lippencott Williams & Wilkins, Baltimore Md. (1999)).

The composition comprising a cupredoxin or variant, derivative or structural equivalent thereof used in the invention may be administered in a variety of ways, including by injection (e.g., intradermal, subcutaneous, intramuscular, intraperitoneal and the like), by inhalation, by topical administration, by suppository, by using a transdermal patch or by mouth. General information on drug delivery systems can be found in Ansel et al., id. In some embodiments, the composition comprising a cupredoxin or variant, derivative or structural equivalent thereof can be formulated and used directly as injectibles, for subcutaneous and intravenous injection, among others. The injectable formulation, in particular, can advantageously be used to treat patients that are appropriate for chemopreventive therapy. The composition comprising a cupredoxin or variant, derivative or structural equivalent thereof can also be taken orally after mixing with protective agents such as polypropylene glycols or similar coating agents.

When administration is by injection, the cupredoxin or variant, derivative or structural equivalent thereof may be formulated in aqueous solutions, specifically in physiologically compatible buffers such as Hanks solution, Ringer's solution, or physiological saline buffer. The solution may contain formulatory agents such as suspending, stabilizing and/or dispersing agents. Alternatively, the cupredoxin or variant, derivative or structural equivalent thereof may be in powder form for constitution with a suitable vehicle, e.g., sterile pyrogen-free water, before use. In some embodiments, the pharmaceutical composition does not comprise an adjuvant or any other substance added to enhance the immune response stimulated by the peptide. In some embodiments, the pharmaceutical composition comprises a substance that inhibits an immune response to the peptide.

When administration is by intravenous fluids, the intravenous fluids for use administering the cupredoxin or variant, derivative or structural equivalent thereof may be composed of crystalloids or colloids. Crystalloids as used herein are

aqueous solutions of mineral salts or other water-soluble molecules. Colloids as used herein contain larger insoluble molecules, such as gelatin. Intravenous fluids may be sterile.

Crystalloid fluids that may be used for intravenous administration include but are not limited to, normal saline (a solution of sodium chloride at 0.9% concentration), Ringer's lactate or Ringer's solution, and a solution of 5% dextrose in water sometimes called D5W, as described in Table 2.

TABLE 2

Composition of Common Crystalloid Solutions				
Solution	Other Name	[Na ⁺]	[Cl ⁻]	[Glucose]
D5W	5% Dextrose	0	0	252
2/3 & 1/3	3.3% Dextrose/ 0.3% saline	51	51	168
Half-normal saline	0.45% NaCl	77	77	0
Normal saline	0.9% NaCl	154	154	0
Ringer's lactate*	Ringer's solution	130	109	0

*Ringer's lactate also has 28 mmol/L lactate, 4 mmol/L K⁺ and 3 mmol/L Ca²⁺.

When administration is by inhalation, the cupredoxin or variant, derivative or structural equivalent thereof may be delivered in the form of an aerosol spray from pressurized packs or a nebulizer with the use of a suitable propellant, e.g., dichlorodifluoromethane, trichlorofluoromethane, carbon dioxide or other suitable gas. In the case of a pressurized aerosol, the dosage unit may be determined by providing a valve to deliver a metered amount. Capsules and cartridges of, e.g., gelatin, for use in an inhaler or insufflator may be formulated containing a powder mix of the proteins and a suitable powder base such as lactose or starch.

When administration is by topical administration, the cupredoxin or variant, derivative or structural equivalent thereof may be formulated as solutions, gels, ointments, creams, jellies, suspensions, and the like, as are well known in the art. In some embodiments, administration is by means of a transdermal patch. When administration is by suppository (e.g., rectal or vaginal), cupredoxin or variants and derivatives thereof compositions may also be formulated in compositions containing conventional suppository bases.

When administration is oral, a cupredoxin or variant, derivative or structural equivalent thereof can be readily formulated by combining the cupredoxin or variant, derivative or structural equivalent thereof with pharmaceutically acceptable carriers well known in the art. A solid carrier, such as mannitol, lactose, magnesium stearate, and the like may be employed; such carriers enable the cupredoxin and variants, derivatives or structural equivalent thereof to be formulated as tablets, pills, dragees, capsules, liquids, gels, syrups, slurries, suspensions and the like, for oral ingestion by a subject to be treated. For oral solid formulations such as, for example, powders, capsules and tablets, suitable excipients include fillers such as sugars, cellulose preparation, granulating agents, and binding agents.

Other convenient carriers, as well-known in the art, also include multivalent carriers, such as bacterial capsular polysaccharide, a dextran or a genetically engineered vector. In addition, sustained-release formulations that include a cupredoxin or variant, derivative or structural equivalent thereof allow for the release of cupredoxin or variant, derivative or structural equivalent thereof over extended periods of time, such that without the sustained release formulation, the cupredoxin or variant, derivative or structural equivalent thereof would be cleared from a subject's system, and/or

degraded by, for example, proteases and simple hydrolysis before eliciting or enhancing a therapeutic effect.

The half-life in the bloodstream of the peptides of the invention can be extended or optimized by several methods well known to those in the art. The peptide variants of the invention may include, but are not limited to, various variants that may increase their stability, specific activity, longevity in the bloodstream, and/or decrease immunogenicity of the cupredoxin, while retaining the ability of the peptide to inhibit the development of premalignant lesions in mammalian cells, tissues and animals. Such variants include, but are not limited to, those which decrease the hydrolysis of the peptide, decrease the deamidation of the peptide, decrease the oxidation, decrease the immunogenicity, increase the structural stability of the peptide or increase the size of the peptide. Such peptides also include circularized peptides (see Monk et al., *BioDrugs* 19(4):261-78, (2005); DeFreest et al., *J. Pept. Res.* 63(5):409-19 (2004)), D,L-peptides (diastereomer), Futaki et al., *J. Biol. Chem.* February 23; 276(8):5836-40 (2001); Papo et al., *Cancer Res.* 64(16):5779-86 (2004); Miller et al., *Biochem. Pharmacol.* 36(1):169-76, (1987)); peptides containing unusual amino acids (see Lee et al., *J. Pept. Res.* 63(2):69-84 (2004)), N- and C-terminal modifications (see Labrie et al., *Clin. Invest. Med.* 13(5):275-8, (1990)), hydrocarbon stapling (see Schafineister et al., *J. Am. Chem. Soc.* 122:5891-5892 (2000); Walenski et al., *Science* 305:1466-1470 (2004)) and PEGylation.

In various embodiments, the pharmaceutical composition includes carriers and excipients (including but not limited to buffers, carbohydrates, mannitol, proteins, polypeptides or amino acids such as glycine, antioxidants, bacteriostats, chelating agents, suspending agents, thickening agents and/or preservatives), water, oils, saline solutions, aqueous dextrose and glycerol solutions, other pharmaceutically acceptable auxiliary substances as required to approximate physiological conditions, such as buffering agents, tonicity adjusting agents, wetting agents and the like. It will be recognized that, while any suitable carrier known to those of ordinary skill in the art may be employed to administer the compositions of this invention, the type of carrier will vary depending on the mode of administration. Compounds may also be encapsulated within liposomes using well-known technology. Biodegradable microspheres may also be employed as carriers for the pharmaceutical compositions of this invention. Suitable biodegradable microspheres are disclosed, for example, in U.S. Pat. Nos. 4,897,268; 5,075,109; 5,928,647; 5,811,128; 5,820,883; 5,853,763; 5,814,344 and 5,942,252.

The pharmaceutical compositions may be sterilized by conventional, well-known sterilization techniques, or may be sterile filtered. The resulting aqueous solutions may be packaged for use as is, or lyophilized, the lyophilized preparation being combined with a sterile solution prior to administration. Administration of Cupredoxin or Variant, Derivative or Structural Equivalent Thereof

The cupredoxin or variant, derivative or structural equivalent thereof can be administered formulated as pharmaceutical compositions and administered by any suitable route, for example, by oral, buccal, inhalation, sublingual, rectal, vaginal, transurethral, nasal, topical, percutaneous, i.e., transdermal or parenteral (including intravenous, intramuscular, subcutaneous and intracoronary) or vitreous administration. The pharmaceutical formulations thereof can be administered in any amount effective to achieve its intended purpose. More specifically, the composition is administered in a therapeutically effective amount. In specific embodiments, the therapeutically effective amount is generally from about 0.01-20 mg/day/kg of body weight.

The compounds comprising cupredoxin or variant, derivative or structural equivalent thereof are useful for the prevention of cancer, alone or in combination with other active agents. The appropriate dosage will, of course, vary depending upon, for example, the compound of cupredoxin or variant, derivative or structural equivalent thereof employed, the host, the mode of administration and the nature and severity of the potential cancer. However, in general, satisfactory results in humans are indicated to be obtained at daily dosages from about 0.01-20 mg/kg of body weight. An indicated daily dosage in humans is in the range from about 0.7 mg to about 1400 mg of a compound of cupredoxin or variant, derivative or structural equivalent thereof conveniently administered, for example, in daily doses, weekly doses, monthly doses, and/or continuous dosing. Daily doses can be in discrete dosages from 1 to 12 times per day. Alternatively, doses can be administered every other day, every third day, every fourth day, every fifth day, every sixth day, every week, and similarly in day increments up to 31 days or over. Alternatively, dosing can be continuous using patches, i.v. administration and the like.

The exact formulation, route of administration, and dosage is determined by the attending physician in view of the patient's condition. Dosage amount and interval can be adjusted individually to provide plasma levels of the active cupredoxin or variant, derivative or structural equivalent thereof which are sufficient to maintain therapeutic effect. Generally, the desired cupredoxin or variant, derivative or structural equivalent thereof is administered in an admixture with a pharmaceutical carrier selected with regard to the intended route of administration and standard pharmaceutical practice.

In one aspect, the cupredoxin or variant, derivative or structural equivalent thereof is delivered as DNA such that the polypeptide is generated in situ. In one embodiment, the DNA is "naked," as described, for example, in Ulmer et al., (Science 259:1745-1749 (1993)) and reviewed by Cohen (Science 259:1691-1692 (1993)). The uptake of naked DNA may be increased by coating the DNA onto a carrier, e.g., biodegradable beads, which are then efficiently transported into the cells. In such methods, the DNA may be present within any of a variety of delivery systems known to those of ordinary skill in the art, including nucleic acid expression systems, bacterial and viral expression systems. Techniques for incorporating DNA into such expression systems are well known to those of ordinary skill in the art. See, e.g., WO90/11092, WO93/24640, WO 93/17706, and U.S. Pat. No. 5,736,524.

Vectors, used to shuttle genetic material from organism to organism, can be divided into two general classes: Cloning vectors are replicating plasmid or phage with regions that are essential for propagation in an appropriate host cell and into which foreign DNA can be inserted; the foreign DNA is replicated and propagated as if it were a component of the vector. An expression vector (such as a plasmid, yeast, or animal virus genome) is used to introduce foreign genetic material into a host cell or tissue in order to transcribe and translate the foreign DNA, such as the DNA of a cupredoxin. In expression vectors, the introduced DNA is operably-linked to elements such as promoters that signal to the host cell to highly transcribe the inserted DNA. Some promoters are exceptionally useful, such as inducible promoters that control gene transcription in response to specific factors. Operably-linking a cupredoxin and variants and derivatives thereof polynucleotide to an inducible promoter can control the expression of the cupredoxin and variants and derivatives thereof in response to specific factors. Examples of classic inducible promoters include those that are responsive to α -interferon, heat shock, heavy metal ions, and steroids such as

glucocorticoids (Kaufman, Methods Enzymol. 185:487-511 (1990)) and tetracycline. Other desirable inducible promoters include those that are not endogenous to the cells in which the construct is being introduced, but, are responsive in those cells when the induction agent is exogenously supplied. In general, useful expression vectors are often plasmids. However, other forms of expression vectors, such as viral vectors (e.g., replication defective retroviruses, adenoviruses and adeno-associated viruses) are contemplated.

Vector choice is dictated by the organism or cells being used and the desired fate of the vector. In general, vectors comprise signal sequences, origins of replication, marker genes, polylinker sites, enhancer elements, promoters, and transcription termination sequences.

Kits Comprising Cupredoxin, or Variant, Derivative or Structural Equivalent Thereof

In one aspect, the invention provides regimens or kits comprising one or more of the following in a package or container: (1) a pharmacologically active composition comprising at least one cupredoxin or variant, derivative or structural equivalent thereof; (2) an additional chemopreventive drug, (3) apparatus to administer the biologically active composition to the patient, such as a syringe, nebulizer etc.

When a kit is supplied, the different components of the composition may be packaged in separate containers, if appropriate, and admixed immediately before use. Such packaging of the components separately may permit long-term storage without losing the active components' functions.

The reagents included in the kits can be supplied in containers of any sort such that the life of the different components are preserved and are not adsorbed or altered by the materials of the container. For example, sealed glass ampules may contain lyophilized cupredoxin and variants, derivatives and structural equivalents thereof, or buffers that have been packaged under a neutral, non-reacting gas, such as nitrogen. Ampules may consist of any suitable material, such as glass, organic polymers, such as polycarbonate, polystyrene, etc., ceramic, metal or any other material typically employed to hold similar reagents. Other examples of suitable containers include simple bottles that may be fabricated from similar substances as ampules, and envelopes, that may comprise foil-lined interiors, such as aluminum or an alloy. Other containers include test tubes, vials, flasks, bottles, syringes, or the like. Containers may have a sterile access port, such as a bottle having a stopper that can be pierced by a hypodermic injection needle. Other containers may have two compartments that are separated by a readily removable membrane that upon removal permits the components to be mixed. Removable membranes may be glass, plastic, rubber, etc.

Kits may also be supplied with instructional materials. Instructions may be printed on paper or other substrate, and/or may be supplied as an electronic-readable medium, such as a floppy disc, CD-ROM, DVD-ROM, Zip disc, videotape, audiotape, flash memory device etc. Detailed instructions may not be physically associated with the kit; instead, a user may be directed to an internet web site specified by the manufacturer or distributor of the kit, or supplied as electronic mail. Modification of Cupredoxin and Variants, Derivatives and Structural Equivalents Thereof

Cupredoxin or variant, derivative or structural equivalents thereof may be chemically modified or genetically altered to produce variants and derivatives as explained above. Such variants and derivatives may be synthesized by standard techniques.

In addition to naturally-occurring allelic variants of cupredoxin, changes can be introduced by mutation into cupredoxin coding sequence that incur alterations in the amino acid sequences of the encoded cupredoxin that do not significantly alter the ability of cupredoxin to inhibit the development of

pre-malignant lesions. A "non-essential" amino acid residue is a residue that can be altered from the wild-type sequences of the cupredoxin without altering pharmacologic activity, whereas an "essential" amino acid residue is required for such pharmacologic activity. For example, amino acid residues that are conserved among the cupredoxins are predicted to be particularly non-amenable to alteration, and thus "essential."

Amino acids for which conservative substitutions that do not change the pharmacologic activity of the polypeptide can be made are well known in the art. Useful conservative substitutions are shown in Table 3, "Preferred substitutions." Conservative substitutions whereby an amino acid of one class is replaced with another amino acid of the same type fall within the scope of the invention so long as the substitution does not materially alter the pharmacologic activity of the compound.

TABLE 3

Preferred substitutions		
Original residue	Exemplary substitutions	Preferred substitutions
Ala (A)	Val, Leu, Ile	Val
Arg (R)	Lys, Gln, Asn	Lys
Asn (N)	Gln, His, Lys, Arg	Gln
Asp (D)	Glu	Glu
Cys (C)	Ser	Ser
Gln (Q)	Asn	Asn
Glu (E)	Asp	Asp
Gly (G)	Pro, Ala	Ala
His (H)	Asn, Gln, Lys, Arg	Arg
Ile (I)	Leu, Val, Met, Ala, Phe, Norleucine	Leu
Leu (L)	Norleucine, Ile, Val, Met, Ala, Phe	Ile
Lys (K)	Arg, Gln, Asn	Arg
Met (M)	Leu, Phe, Ile	Leu
Phe (F)	Leu, Val, Ile, Ala, Tyr	Leu
Pro (P)	Ala	Ala
Ser (S)	Thr	Thr
Thr (T)	Ser	Ser
Trp (W)	Tyr, Phe	Tyr
Tyr (Y)	Trp, Phe, Thr, Ser	Phe
Val (V)	Ile, Leu, Met, Phe, Ala, Norleucine	Leu

Non-conservative substitutions that affect (1) the structure of the polypeptide backbone, such as a β -sheet or α -helical conformation, (2) the charge, (3) hydrophobicity, or (4) the bulk of the side chain of the target site can modify the pharmacologic activity. Residues are divided into groups based on common side-chain properties as denoted in Table 4. Non-conservative substitutions entail exchanging a member of one of these classes for another class. Substitutions may be introduced into conservative substitution sites or more specifically into non-conserved sites.

TABLE 4

Amino acid classes	
Class	Amino acids
hydrophobic	Norleucine, Met, Ala, Val, Leu, Ile
neutral hydrophilic	Cys, Ser, Thr
acidic	Asp, Glu
basic	Asn, Gln, His, Lys, Arg
disrupt chain conformation	Gly, Pro
aromatic	Trp, Tyr, Phe

The variant polypeptides can be made using methods known in the art such as oligonucleotide-mediated (site-di-

rected) mutagenesis, alanine scanning, and PCR mutagenesis. Site-directed mutagenesis (Carter, Biochem J. 237:1-7 (1986); Zoller and Smith, Methods Enzymol. 154:329-350 (1987)), cassette mutagenesis, restriction selection mutagenesis (Wells et al., Gene 34:315-323 (1985)) or other known techniques can be performed on the cloned DNA to produce the cupredoxin variant DNA.

Known mutations of cupredoxins can also be used to create variant cupredoxin to be used in the methods of the invention. For example, the C112D and M44KM64E mutants of azurin are known to have cytotoxic and growth arresting activity that is different from the native azurin, and such altered activity can be useful in the treatment methods of the present invention.

A more complete understanding of the present invention can be obtained by reference to the following specific Examples. The Examples are described solely for purposes of illustration and are not intended to limit the scope of the invention. Changes in form and substitution of equivalents are contemplated as circumstances may suggest or render expedient. Although specific terms have been employed herein, such terms are intended in a descriptive sense and not for purposes of limitations. Modifications and variations of the invention as hereinbefore set forth can be made without departing from the spirit and scope thereof.

EXAMPLES

Example 1

Effect of Peptide P-28 on DMBA-Induced Mammary Lesions in the MMOC Model

The mouse mammary gland organ culture (MMOC) model allows evaluating efficacy of potentially chemopreventive agents against development of mammary alveolar lesions (MAL) or mammary ductal lesions (MDL) in response to DMBA. DMBA under appropriate incubation conditions form either MAL or MDL based on the hormonal milieu in the medium. Hawthorne et al., Pharmaceutical Biology 40: 70-74 (2002); Mehta et al., J. Natl. Cancer Inst. 93: 1103-1106 (2001). Estrogen and progesterone-treated glands in culture develop ductal lesions whereas aldosterone and hydrocortisone-treated glands form estrogen and progesterone-independent alveolar lesions. Mammary glands not exposed to a carcinogen or chemopreventive agent, undergo structural regression in the absence of growth-promoting hormones, whereas treatment with DMBA for the 24-hr period between days 3 and 4 prevents the regression of structures caused by deprivation of hormones. It is assumed that this is because the glands have lost normal hormonal responsiveness and now have altered their course of development. Generating mammary adenocarcinoma by transplanting transformed cells into syngeneic mice has proved the premalignant pre-neoplastic nature of these unrepressed areas.

The thoracic pair of mammary glands was excised aseptically from each Balb/c mouse, and the glands were divided into several groups. The effects of p28 were evaluated at 4 different dilutions in the medium. Carcinogen treated glands without the test agent served as a measure to determine percent incidence in the absence of a chemopreventive agent. An additional control was included to serve as a positive control for chemoprevention. Azurin was included in the medium at 50 μ g/ml concentration. For alveolar lesions (MAL) stained glands were evaluated for the incidence of lesions (glands containing any lesions as compared to total number of glands in a given treatment group). For the ductal lesions (MDL)

similar protocol was adapted, however, as indicated below in the methods section the hormonal combination is different for alveolar and ductal lesions. The glands were fixed in formalin and then processed for histopathology. The sections are stained with eosin and hematoxyline and evaluated under microscope. Here the multiplicity of ductal lesions between the control and the treatment groups are compared.

Organ Culture Procedure.

The experimental animals used for the studies were young, virgin BALB/c female mice 3 to 4 weeks of age obtained from Charles River, Wilmington, Mass. The mice were treated daily by subcutaneous injections with 1 μ g estradiol-17 β +1 mg progesterone for 9 days. This treatment is a prerequisite inasmuch as animals not pretreated with steroids fail to respond to hormones in vitro. The entire culture procedure is described in detail. Jang et al., *Science* 275:218-220 (1997); Mehta, *Eu. J. Cancer* 36:1275-1282 (2000); Mehta et al., *J. Natl. Cancer Inst.* 89:212-219 (1997); Mehta et al., *J. Natl. Cancer Inst.* 93:1103-1106 (2001).

Briefly, the animals were killed by cervical dislocation, and the thoracic pair of mammary glands were dissected out on silk rafts and incubated for 10 days in serum free Waymouth MB752/1 medium (5-glands/5 ml/dish). The medium was supplemented with glutamine, antibiotics (penicillin and streptomycin 100 units/ml medium) and growth-promoting hormones, 5 μ g insulin (I), 5 μ g prolactin (P), 1 μ g aldosterone (A) and 1 μ g hydrocortisone (H) per ml of medium for the protocol to induce mammary alveolar lesions (MAL). For induction of ductal lesions (MDL), the medium contained 5 μ g/ml, 5 μ g/ml P, 0.001 μ g/ml estradiol 17 β and 1 μ g/ml progesterone (Pg). Mehta et al., *J. Natl. Cancer Inst.* 93:1103-1106 (2001). The carcinogen, DMBA (2 μ g/ml) was added to the medium between days 3 and 4. For the present study, DMBA was dissolved in DMSO at a final concentration of 4 mg/ml, and 50 μ g I was added to 100 ml medium resulting in 2 μ g/ml final concentrations. The control dishes contained DMSO as vehicle.

On day 4, DMBA is removed from the medium by rinsing the glands in fresh medium and transferring them to new dishes containing fresh medium without DMBA. After 10 days of incubation, the glands were maintained for another 14 days in the medium containing only I (5 μ g/ml). During the entire culture period, the glands were maintained at 37° C. under 95% O₂ and 5% CO₂ environment. The chemopreventive agent was included in the medium during the first ten days of growth-promoting phase. The test peptide p28 was evaluated at 4 concentrations ranging from 12.5 μ g/ml to 100 μ g/ml. Azurin was evaluated at 50 μ g/ml in the medium. The peptide was dissolved in sterile water and filtered prior to use. The medium was changed three times per week (Monday, Wednesday and Friday). At the end of the exposure, the glands were fixed in formalin.

Results were analyzed by Chi-square analysis and Fisher's Exact Test.

Morphometric Analysis of MAL.

For examination of MAL, the glands were stained in alum carmine, and evaluated for the presence of the lesions. The glands were scored for the presence or absence of mammary lesions, severity of lesions per gland, and toxicity of the agent. The glands stored in xylene were evaluated for the presence or absence, incidence, and severity of mammary lesions for each

gland under a dissecting microscope. Mammary glands were scored as positive or negative for mammary lesions, and the percent incidence was determined as a ratio of glands exhibiting lesions and the total number of glands in that group. Dilution of ducts or disintegration of mammary structure because of treatment with chemopreventive agent was considered a toxic effect. The data were subjected to statistical analysis for the incidence to determine the effectiveness of the potential chemopreventive agents.

FIG. 1A shows a representative photograph of alveolar lesions in a DMBA treated gland and its comparison with a gland that was treated with DMBA along with a chemopreventive agent. The effects of p28 on the development of alveolar lesion are shown in FIGS. 1B-1G and summarized in FIG. 2. The peptide p28 inhibited MAL formation by 67% at 25 μ g/ml concentration. Increasing concentration further up to 100 μ g/ml did not enhance the efficacy of the peptide. The comparison of the peptide with azurin indicated that p28 was as effective as azurin for MAL development. Azurin at 50 μ g/ml concentration resulted in a 67% inhibition. Statistical analyses indicated that the effect of p28 was statistically significant compared to DMBA control at concentrations greater than 12.5 μ g/ml ($p < 0.01$, Fisher's Exact Test; Chi Square analysis).

Histopathological Evaluation of MDL.

For MDL, the glands were processed for histopathological evaluations. The glands were sectioned longitudinally into 5-micron sections and stained with eosin hematoxyline. The longitudinal section of each gland was divided into several fields and each field was evaluated for ductal lesions. Mehta et al., *J. Natl. Cancer Inst.* 93:1103-1106 (2001). Briefly, the entire gland is evaluated under the scope; smaller glands will have fewer total fields as compared to larger glands. Thus, each gland will have variable number of fields. Often the number of sections through the ducts also varies greatly from gland to gland. This results in the variable number from group to group. Fields containing ductal hyperplasia or atypia were determined and were compared with total number of field evaluated for each gland. No discrimination is made between the hyperplasia or atypia and severely occluded glands. Any field containing any of these histological patterns was considered positive for the lesion. The treatment groups were compared with the controls for the severity and percent inhibition was calculated.

FIG. 3 shows a representative ductal lesion. DMBA induces ductal lesions varying from hyperplasia, atypia to complete occlusion of the ducts. A ratio of ductal lesions/total number of ductal sections was determined. Again, 12.5 μ g/ml concentration of p28 suppressed only 15% of the MDL formation. However, at 25 μ g/ml there was a significant inhibition of the lesions comparable to that observed with 50 μ g/ml azurin. The efficacy of p28 at concentrations greater than 12.5 μ g/ml was statistically significant ($p < 0.01$, Fishers Exact Test). These results are summarized in FIG. 4. Often effects of chemopreventive agents can be differentiated between the MAL and MDL. For example tamoxifen inhibited the development of MDL but not MAL. It is interesting to note that azurin and p28 inhibited both estrogen and progesterone-dependent ductal lesions as well as independent alveolar lesions.

This example indicates that both p28 and azurin can prevent the development of precancerous lesions in breast tissue. Thus, p28 and azurin may be used as chemopreventive agents in mammalian patients.

SEQUENCE LISTING

<160> NUMBER OF SEQ ID NOS: 26

<210> SEQ ID NO 1

<211> LENGTH: 128

<212> TYPE: PRT

<213> ORGANISM: *Pseudomonas aeruginosa*

<400> SEQUENCE: 1

```

Ala Glu Cys Ser Val Asp Ile Gln Gly Asn Asp Gln Met Gln Phe Asn
1          5          10          15
Thr Asn Ala Ile Thr Val Asp Lys Ser Cys Lys Gln Phe Thr Val Asn
          20          25          30
Leu Ser His Pro Gly Asn Leu Pro Lys Asn Val Met Gly His Asn Trp
          35          40          45
Val Leu Ser Thr Ala Ala Asp Met Gln Gly Val Val Thr Asp Gly Met
          50          55          60
Ala Ser Gly Leu Asp Lys Asp Tyr Leu Lys Pro Asp Asp Ser Arg Val
65          70          75          80
Ile Ala His Thr Lys Leu Ile Gly Ser Gly Glu Lys Asp Ser Val Thr
          85          90          95
Phe Asp Val Ser Lys Leu Lys Glu Gly Glu Gln Tyr Met Phe Phe Cys
          100          105          110
Thr Phe Pro Gly His Ser Ala Leu Met Lys Gly Thr Leu Thr Leu Lys
          115          120          125

```

<210> SEQ ID NO 2

<211> LENGTH: 28

<212> TYPE: PRT

<213> ORGANISM: *Pseudomonas aeruginosa*

<400> SEQUENCE: 2

```

Leu Ser Thr Ala Ala Asp Met Gln Gly Val Val Thr Asp Gly Met Ala
1          5          10          15
Ser Gly Leu Asp Lys Asp Tyr Leu Lys Pro Asp Asp
          20          25

```

<210> SEQ ID NO 3

<211> LENGTH: 105

<212> TYPE: PRT

<213> ORGANISM: *Phormidium laminosum*

<400> SEQUENCE: 3

```

Glu Thr Phe Thr Val Lys Met Gly Ala Asp Ser Gly Leu Leu Gln Phe
1          5          10          15
Glu Pro Ala Asn Val Thr Val His Pro Gly Asp Thr Val Lys Trp Val
          20          25          30
Asn Asn Lys Leu Pro Pro His Asn Ile Leu Phe Asp Asp Lys Gln Val
          35          40          45
Pro Gly Ala Ser Lys Glu Leu Ala Asp Lys Leu Ser His Ser Gln Leu
          50          55          60
Met Phe Ser Pro Gly Glu Ser Tyr Glu Ile Thr Phe Ser Ser Asp Phe
65          70          75          80
Pro Ala Gly Thr Tyr Thr Tyr Tyr Cys Ala Pro His Arg Gly Ala Gly
          85          90          95
Met Val Gly Lys Ile Thr Val Glu Gly
          100          105

```

<210> SEQ ID NO 4

-continued

```

<211> LENGTH: 155
<212> TYPE: PRT
<213> ORGANISM: Thiobacillus ferrooxidans

<400> SEQUENCE: 4

Gly Thr Leu Asp Thr Thr Trp Lys Glu Ala Thr Leu Pro Gln Val Lys
1          5          10          15

Ala Met Leu Glu Lys Asp Thr Gly Lys Val Ser Gly Asp Thr Val Thr
20          25          30

Tyr Ser Gly Lys Thr Val His Val Val Ala Ala Val Leu Pro Gly
35          40          45

Phe Pro Phe Pro Ser Phe Glu Val His Asp Lys Lys Asn Pro Thr Leu
50          55          60

Glu Ile Pro Ala Gly Ala Thr Val Asp Val Thr Phe Ile Asn Thr Asn
65          70          75          80

Lys Gly Phe Gly His Ser Phe Asp Ile Thr Lys Lys Gly Pro Pro Tyr
85          90          95

Ala Val Met Pro Val Ile Asp Pro Ile Val Ala Gly Thr Gly Phe Ser
100         105         110

Pro Val Pro Lys Asp Gly Lys Phe Gly Tyr Thr Asp Phe Thr Trp His
115         120         125

Pro Thr Ala Gly Thr Tyr Tyr Tyr Val Cys Gln Ile Pro Gly His Ala
130         135         140

Ala Thr Gly Met Phe Gly Lys Ile Val Val Lys
145         150         155

```

```

<210> SEQ ID NO 5
<211> LENGTH: 124
<212> TYPE: PRT
<213> ORGANISM: Achromobacter cycloclastes

<400> SEQUENCE: 5

Ala Asp Phe Glu Val His Met Leu Asn Lys Gly Lys Asp Gly Ala Met
1          5          10          15

Val Phe Glu Pro Ala Ser Leu Lys Val Ala Pro Gly Asp Thr Val Thr
20          25          30

Phe Ile Pro Thr Asp Lys Gly His Asn Val Glu Thr Ile Lys Gly Met
35          40          45

Ile Pro Asp Gly Ala Glu Ala Phe Lys Ser Lys Ile Asn Glu Asn Tyr
50          55          60

Lys Val Thr Phe Thr Ala Pro Gly Val Tyr Gly Val Lys Cys Thr Pro
65          70          75          80

His Tyr Gly Met Gly Met Val Gly Val Val Gln Val Gly Asp Ala Pro
85          90          95

Ala Asn Leu Glu Ala Val Lys Gly Ala Lys Asn Pro Lys Lys Ala Gln
100         105         110

Glu Arg Leu Asp Ala Ala Leu Ala Leu Gly Asn
115         120

```

```

<210> SEQ ID NO 6
<211> LENGTH: 128
<212> TYPE: PRT
<213> ORGANISM: Alcaligenes faecalis

<400> SEQUENCE: 6

Ala Cys Asp Val Ser Ile Glu Gly Asn Asp Ser Met Gln Phe Asn Thr
1          5          10          15

```

-continued

Lys Ser Ile Val Val Asp Lys Thr Cys Lys Glu Phe Thr Ile Asn Leu
 20 25 30
 Lys His Thr Gly Lys Leu Pro Lys Ala Ala Met Gly His Asn Val Val
 35 40 45
 Val Ser Lys Lys Ser Asp Glu Ser Ala Val Ala Thr Asp Gly Met Lys
 50 55 60
 Ala Gly Leu Asn Asn Asp Tyr Val Lys Ala Gly Asp Glu Arg Val Ile
 65 70 75 80
 Ala His Thr Ser Val Ile Gly Gly Gly Glu Thr Asp Ser Val Thr Phe
 85 90 95
 Asp Val Ser Lys Leu Lys Glu Gly Glu Asp Tyr Ala Phe Phe Cys Ser
 100 105 110
 Phe Pro Gly His Trp Ser Ile Met Lys Gly Thr Ile Glu Leu Gly Ser
 115 120 125

<210> SEQ ID NO 7
 <211> LENGTH: 129
 <212> TYPE: PRT
 <213> ORGANISM: *Achromobacter xylosoxidans*

<400> SEQUENCE: 7

Ala Gln Cys Glu Ala Thr Ile Glu Ser Asn Asp Ala Met Gln Tyr Asn
 1 5 10 15
 Leu Lys Glu Met Val Val Asp Lys Ser Cys Lys Gln Phe Thr Val His
 20 25 30
 Leu Lys His Val Gly Lys Met Ala Lys Val Ala Met Gly His Asn Trp
 35 40 45
 Val Leu Thr Lys Glu Ala Asp Lys Gln Gly Val Ala Thr Asp Gly Met
 50 55 60
 Asn Ala Gly Leu Ala Gln Asp Tyr Val Lys Ala Gly Asp Thr Arg Val
 65 70 75 80
 Ile Ala His Thr Lys Val Ile Gly Gly Gly Glu Ser Asp Ser Val Thr
 85 90 95
 Phe Asp Val Ser Lys Leu Thr Pro Gly Glu Ala Tyr Ala Tyr Phe Cys
 100 105 110
 Ser Phe Pro Gly His Trp Ala Met Met Lys Gly Thr Leu Lys Leu Ser
 115 120 125

Asn

<210> SEQ ID NO 8
 <211> LENGTH: 129
 <212> TYPE: PRT
 <213> ORGANISM: *Bordetella bronchiseptica*

<400> SEQUENCE: 8

Ala Glu Cys Ser Val Asp Ile Ala Gly Thr Asp Gln Met Gln Phe Asp
 1 5 10 15
 Lys Lys Ala Ile Glu Val Ser Lys Ser Cys Lys Gln Phe Thr Val Asn
 20 25 30
 Leu Lys His Thr Gly Lys Leu Pro Arg Asn Val Met Gly His Asn Trp
 35 40 45
 Val Leu Thr Lys Thr Ala Asp Met Gln Ala Val Glu Lys Asp Gly Ile
 50 55 60
 Ala Ala Gly Leu Asp Asn Gln Tyr Leu Lys Ala Gly Asp Thr Arg Val
 65 70 75 80
 Leu Ala His Thr Lys Val Leu Gly Gly Gly Glu Ser Asp Ser Val Thr

-continued

	85		90		95										
Phe	Asp	Val	Ala	Lys	Leu	Ala	Ala	Gly	Asp	Asp	Tyr	Thr	Phe	Phe	Cys
	100							105					110		
Ser	Phe	Pro	Gly	His	Gly	Ala	Leu	Met	Lys	Gly	Thr	Leu	Lys	Leu	Val
	115						120					125			

Asp

<210> SEQ ID NO 9
 <211> LENGTH: 129
 <212> TYPE: PRT
 <213> ORGANISM: Methylobacterium sp.

<400> SEQUENCE: 9

Ala	Ser	Cys	Glu	Thr	Thr	Val	Thr	Ser	Gly	Asp	Thr	Met	Thr	Tyr	Ser
1			5						10					15	
Thr	Arg	Ser	Ile	Ser	Val	Pro	Ala	Ser	Cys	Ala	Glu	Phe	Thr	Val	Asn
	20						25						30		
Phe	Glu	His	Lys	Gly	His	Met	Pro	Lys	Thr	Gly	Met	Gly	His	Asn	Trp
	35					40						45			
Val	Leu	Ala	Lys	Ser	Ala	Asp	Val	Gly	Asp	Val	Ala	Lys	Glu	Gly	Ala
	50					55					60				
His	Ala	Gly	Ala	Asp	Asn	Asn	Phe	Val	Thr	Pro	Gly	Asp	Lys	Arg	Val
65				70					75					80	
Ile	Ala	Phe	Thr	Pro	Ile	Ile	Gly	Gly	Gly	Glu	Lys	Thr	Ser	Val	Lys
	85						90							95	
Phe	Lys	Val	Ser	Ala	Leu	Ser	Lys	Asp	Glu	Ala	Tyr	Thr	Tyr	Phe	Cys
	100						105						110		
Ser	Tyr	Pro	Gly	His	Phe	Ser	Met	Met	Arg	Gly	Thr	Leu	Lys	Leu	Glu
	115						120					125			

Glu

<210> SEQ ID NO 10
 <211> LENGTH: 166
 <212> TYPE: PRT
 <213> ORGANISM: Neisseria meningitidis

<400> SEQUENCE: 10

Cys	Ser	Gln	Glu	Pro	Ala	Ala	Pro	Ala	Ala	Glu	Ala	Thr	Pro	Ala	Ala
1			5						10					15	
Glu	Ala	Pro	Ala	Ser	Glu	Ala	Pro	Ala	Ala	Glu	Ala	Ala	Pro	Ala	Asp
	20						25						30		
Ala	Ala	Glu	Ala	Pro	Ala	Ala	Gly	Asn	Cys	Ala	Ala	Thr	Val	Glu	Ser
	35					40						45			
Asn	Asp	Asn	Met	Gln	Phe	Asn	Thr	Lys	Asp	Ile	Gln	Val	Ser	Lys	Ala
	50					55				60					
Cys	Lys	Glu	Phe	Thr	Ile	Thr	Leu	Lys	His	Thr	Gly	Thr	Gln	Pro	Lys
65				70					75					80	
Thr	Ser	Met	Gly	His	Asn	Ile	Val	Ile	Gly	Lys	Thr	Glu	Asp	Met	Asp
	85					90							95		
Gly	Ile	Phe	Lys	Asp	Gly	Val	Gly	Ala	Ala	Asp	Thr	Asp	Tyr	Val	Lys
	100						105						110		
Pro	Asp	Asp	Ala	Arg	Val	Val	Ala	His	Thr	Lys	Leu	Ile	Gly	Gly	Gly
	115						120					125			
Glu	Glu	Ser	Ser	Leu	Thr	Leu	Asp	Pro	Ala	Lys	Leu	Ala	Asp	Gly	Glu
	130					135						140			

-continued

Tyr Lys Phe Ala Cys Thr Phe Pro Gly His Gly Ala Leu Met Asn Gly
145 150 155 160

Lys Val Thr Leu Val Asp
165

<210> SEQ ID NO 11
<211> LENGTH: 128
<212> TYPE: PRT
<213> ORGANISM: *Pseudomonas fluorescens*

<400> SEQUENCE: 11

Ala Glu Cys Lys Thr Thr Ile Asp Ser Thr Asp Gln Met Ser Phe Asn
1 5 10 15

Thr Lys Ala Ile Glu Ile Asp Lys Ala Cys Lys Thr Phe Thr Val Glu
20 25 30

Leu Thr His Ser Gly Ser Leu Pro Lys Asn Val Met Gly His Asn Leu
35 40 45

Val Ile Ser Lys Gln Ala Asp Met Gln Pro Ile Ala Thr Asp Gly Leu
50 55 60

Ser Ala Gly Ile Asp Lys Asn Tyr Leu Lys Glu Gly Asp Thr Arg Val
65 70 75 80

Ile Ala His Thr Lys Val Ile Gly Ala Gly Glu Lys Asp Ser Leu Thr
85 90 95

Ile Asp Val Ser Lys Leu Asn Ala Ala Glu Lys Tyr Gly Phe Phe Cys
100 105 110

Ser Phe Pro Gly His Ile Ser Met Met Lys Gly Thr Val Thr Leu Lys
115 120 125

<210> SEQ ID NO 12
<211> LENGTH: 128
<212> TYPE: PRT
<213> ORGANISM: *Pseudomonas chlororaphis*

<400> SEQUENCE: 12

Ala Glu Cys Lys Val Asp Val Asp Ser Thr Asp Gln Met Ser Phe Asn
1 5 10 15

Thr Lys Glu Ile Thr Ile Asp Lys Ser Cys Lys Thr Phe Thr Val Asn
20 25 30

Leu Thr His Ser Gly Ser Leu Pro Lys Asn Val Met Gly His Asn Trp
35 40 45

Val Leu Ser Lys Ser Ala Asp Met Ala Gly Ile Ala Thr Asp Gly Met
50 55 60

Ala Ala Gly Ile Asp Lys Asp Tyr Leu Lys Pro Gly Asp Ser Arg Val
65 70 75 80

Ile Ala His Thr Lys Ile Ile Gly Ser Gly Glu Lys Asp Ser Val Thr
85 90 95

Phe Asp Val Ser Lys Leu Thr Ala Gly Glu Ser Tyr Glu Phe Phe Cys
100 105 110

Ser Phe Pro Gly His Asn Ser Met Met Lys Gly Ala Val Val Leu Lys
115 120 125

<210> SEQ ID NO 13
<211> LENGTH: 129
<212> TYPE: PRT
<213> ORGANISM: *Xylella fastidiosa*

<400> SEQUENCE: 13

Lys Thr Cys Ala Val Thr Ile Ser Ala Asn Asp Gln Met Lys Phe Asp

-continued

1	5	10	15
Gln Asn Thr	Ile Lys Ile Ala Ala	Glu Cys Thr His Val	Asn Leu Thr
	20	25	30
Leu Thr His	Thr Gly Lys Lys Ser	Ala Arg Val Met Gly	His Asn Trp
	35	40	45
Val Leu Thr	Lys Thr Thr Asp Met	Gln Ala Val Ala	Leu Ala Gly Leu
	50	55	60
His Ala Thr	Leu Ala Asp Asn Tyr	Val Pro Lys Ala	Asp Pro Arg Val
	65	70	75
Ile Ala His	Thr Ala Ile Ile Gly	Gly Gly Glu Arg	Thr Ser Ile Thr
	85	90	95
Phe Pro Thr	Asn Thr Leu Ser Lys	Asn Val Ser Tyr	Thr Phe Phe Cys
	100	105	110
Ser Phe Pro	Gly His Trp Ala Leu	Met Lys Gly Thr	Leu Asn Phe Gly
	115	120	125

Gly

<210> SEQ ID NO 14
 <211> LENGTH: 138
 <212> TYPE: PRT
 <213> ORGANISM: Cucumis sativus

<400> SEQUENCE: 14

Met Gln Ser	Thr Val His Ile Val	Gly Asp Asn Thr	Gly Trp Ser Val
1	5	10	15
Pro Ser Ser	Pro Asn Phe Tyr Ser	Gln Trp Ala Ala	Gly Lys Thr Phe
	20	25	30
Arg Val Gly	Asp Ser Leu Gln Phe	Asn Phe Pro Ala	Asn Ala His Asn
	35	40	45
Val His Glu	Met Glu Thr Lys Gln	Ser Phe Asp Ala	Cys Asn Phe Val
	50	55	60
Asn Ser Asp	Asn Asp Val Glu Arg	Thr Ser Pro Val	Ile Glu Arg Leu
	65	70	75
Asp Glu Leu	Gly Met His Tyr Phe	Val Cys Thr Val	Gly Thr His Cys
	85	90	95
Ser Asn Gly	Gln Lys Leu Ser Ile	Asn Val Val Ala	Ala Asn Ala Thr
	100	105	110
Val Ser Met	Pro Pro Pro Ser	Ser Ser Pro Pro	Ser Val Met Pro
	115	120	125
Pro Pro Val	Met Pro Pro Pro	Ser Ser Pro Ser	
	130	135	

<210> SEQ ID NO 15
 <211> LENGTH: 162
 <212> TYPE: PRT
 <213> ORGANISM: Chloroflexus aurantiacus

<400> SEQUENCE: 15

Met Lys Ile	Thr Leu Arg Met Met	Val Leu Ala Val	Leu Thr Ala Met
1	5	10	15
Ala Met Val	Leu Ala Ala Cys Gly	Gly Gly Gly Ser	Ser Gly Gly Ser
	20	25	30
Thr Gly Gly	Gly Ser Gly Ser	Gly Pro Val Thr	Ile Glu Ile Gly Ser
	35	40	45
Lys Gly Glu	Glu Leu Ala Phe Asp	Lys Thr Glu Leu	Thr Val Ser Ala
	50	55	60

-continued

Gly Gln Thr Val Thr Ile Arg Phe Lys Asn Asn Ser Ala Val Gln Gln
65 70 75 80

His Asn Trp Ile Leu Val Lys Gly Gly Glu Ala Glu Ala Ala Asn Ile
85 90 95

Ala Asn Ala Gly Leu Ser Ala Gly Pro Ala Ala Asn Tyr Leu Pro Ala
100 105 110

Asp Lys Ser Asn Ile Ile Ala Glu Ser Pro Leu Ala Asn Gly Asn Glu
115 120 125

Thr Val Glu Val Thr Phe Thr Ala Pro Ala Ala Gly Thr Tyr Leu Tyr
130 135 140

Ile Cys Thr Val Pro Gly His Tyr Pro Leu Met Gln Gly Lys Leu Val
145 150 155 160

Val Asn

<210> SEQ ID NO 16
<211> LENGTH: 140
<212> TYPE: PRT
<213> ORGANISM: Chloroflexus aurantiacus

<400> SEQUENCE: 16

Ala Ala Asn Ala Pro Gly Gly Ser Asn Val Val Asn Glu Thr Pro Ala
1 5 10 15

Gln Thr Val Glu Val Arg Ala Ala Pro Asp Ala Leu Ala Phe Ala Gln
20 25 30

Thr Ser Leu Ser Leu Pro Ala Asn Thr Val Val Arg Leu Asp Phe Val
35 40 45

Asn Gln Asn Asn Leu Gly Val Gln His Asn Trp Val Leu Val Asn Gly
50 55 60

Gly Asp Asp Val Ala Ala Ala Val Asn Thr Ala Ala Gln Asn Asn Ala
65 70 75 80

Asp Ala Leu Phe Val Pro Pro Pro Asp Thr Pro Asn Ala Leu Ala Trp
85 90 95

Thr Ala Met Leu Asn Ala Gly Glu Ser Gly Ser Val Thr Phe Arg Thr
100 105 110

Pro Ala Pro Gly Thr Tyr Leu Tyr Ile Cys Thr Phe Pro Gly His Tyr
115 120 125

Leu Ala Gly Met Lys Gly Thr Leu Thr Val Thr Pro
130 135 140

<210> SEQ ID NO 17
<211> LENGTH: 96
<212> TYPE: PRT
<213> ORGANISM: Cucumis sativus

<400> SEQUENCE: 17

Ala Val Tyr Val Val Gly Gly Ser Gly Gly Trp Thr Phe Asn Thr Glu
1 5 10 15

Ser Trp Pro Lys Gly Lys Arg Phe Arg Ala Gly Asp Ile Leu Leu Phe
20 25 30

Asn Tyr Asn Pro Ser Met His Asn Val Val Val Val Asn Gln Gly Gly
35 40 45

Phe Ser Thr Cys Asn Thr Pro Ala Gly Ala Lys Val Tyr Thr Ser Gly
50 55 60

Arg Asp Gln Ile Lys Leu Pro Lys Gly Gln Ser Tyr Phe Ile Cys Asn
65 70 75 80

-continued

Phe Pro Gly His Cys Gln Ser Gly Met Lys Ile Ala Val Asn Ala Leu
85 90 95

<210> SEQ ID NO 18
 <211> LENGTH: 166
 <212> TYPE: PRT
 <213> ORGANISM: Neisseria gonorrhoeae

<400> SEQUENCE: 18

Cys Ser Gln Glu Pro Ala Ala Pro Ala Ala Glu Ala Thr Pro Ala Gly
1 5 10 15
 Glu Ala Pro Ala Ser Glu Ala Pro Ala Ala Glu Ala Ala Pro Ala Asp
20 25 30
 Ala Ala Glu Ala Pro Ala Ala Gly Asn Cys Ala Ala Thr Val Glu Ser
35 40 45
 Asn Asp Asn Met Gln Phe Asn Thr Lys Asp Ile Gln Val Ser Lys Ala
50 55 60
 Cys Lys Glu Phe Thr Ile Thr Leu Lys His Thr Gly Thr Gln Pro Lys
65 70 75 80
 Ala Ser Met Gly His Asn Leu Val Ile Ala Lys Ala Glu Asp Met Asp
85 90 95
 Gly Val Phe Lys Asp Gly Val Gly Ala Ala Asp Thr Asp Tyr Val Lys
100 105 110
 Pro Asp Asp Ala Arg Val Val Ala His Thr Lys Leu Ile Gly Gly Gly
115 120 125
 Glu Glu Ser Ser Leu Thr Leu Asp Pro Ala Lys Leu Ala Asp Gly Asp
130 135 140
 Tyr Lys Phe Ala Cys Thr Phe Pro Gly His Gly Ala Leu Met Asn Gly
145 150 155 160
 Lys Val Thr Leu Val Asp
165

<210> SEQ ID NO 19
 <211> LENGTH: 150
 <212> TYPE: PRT
 <213> ORGANISM: Vibrio parahaemolyticus

<400> SEQUENCE: 19

Met Ser Leu Arg Ile Leu Ala Ala Thr Leu Ala Leu Ala Gly Leu Ser
1 5 10 15
 Phe Gly Ala Gln Ala Ser Ala Glu Cys Glu Val Ser Ile Asp Ala Asn
20 25 30
 Asp Met Met Gln Phe Ser Thr Lys Thr Leu Ser Val Pro Ala Thr Cys
35 40 45
 Lys Glu Val Thr Leu Thr Leu Asn His Thr Gly Lys Met Pro Ala Gln
50 55 60
 Ser Met Gly His Asn Val Val Ile Ala Asp Thr Ala Asn Ile Gln Ala
65 70 75 80
 Val Gly Thr Asp Gly Met Ser Ala Gly Ala Asp Asn Ser Tyr Val Lys
85 90 95
 Pro Asp Asp Glu Arg Val Tyr Ala His Thr Lys Val Val Gly Gly Gly
100 105 110
 Glu Ser Thr Ser Ile Thr Phe Ser Thr Glu Lys Met Thr Ala Gly Gly
115 120 125
 Asp Tyr Ser Phe Phe Cys Ser Phe Pro Gly His Trp Ala Ile Met Gln
130 135 140

-continued

Gly Lys Phe Glu Phe Lys
145 150

<210> SEQ ID NO 20
<211> LENGTH: 33
<212> TYPE: PRT
<213> ORGANISM: Chloroflexus aurantiacus

<400> SEQUENCE: 20

His Asn Trp Val Leu Val Asn Gly Gly Asp Asp Val Ala Ala Ala Val
1 5 10 15

Asn Thr Ala Ala Gln Asn Asn Ala Asp Ala Leu Phe Val Pro Pro Pro
20 25 30

Asp

<210> SEQ ID NO 21
<211> LENGTH: 27
<212> TYPE: PRT
<213> ORGANISM: Pseudomonas syringae

<400> SEQUENCE: 21

Ser Lys Lys Ala Asp Ala Ser Ala Ile Thr Thr Asp Gly Met Ser Val
1 5 10 15

Gly Ile Asp Lys Asp Tyr Val Lys Pro Asp Asp
20 25

<210> SEQ ID NO 22
<211> LENGTH: 27
<212> TYPE: PRT
<213> ORGANISM: Neisseria meningitidis

<400> SEQUENCE: 22

Ile Gly Lys Thr Glu Asp Met Asp Gly Ile Phe Lys Asp Gly Val Gly
1 5 10 15

Ala Ala Asp Thr Asp Tyr Val Lys Pro Asp Asp
20 25

<210> SEQ ID NO 23
<211> LENGTH: 27
<212> TYPE: PRT
<213> ORGANISM: Vibrio parahaemolyticus

<400> SEQUENCE: 23

Ala Asp Thr Ala Asn Ile Gln Ala Val Gly Thr Asp Gly Met Ser Ala
1 5 10 15

Gly Ala Asp Asn Ser Tyr Val Lys Pro Asp Asp
20 25

<210> SEQ ID NO 24
<211> LENGTH: 27
<212> TYPE: PRT
<213> ORGANISM: Bordetella bronchiseptica

<400> SEQUENCE: 24

Thr Lys Thr Ala Asp Met Gln Ala Val Glu Lys Asp Gly Ile Ala Ala
1 5 10 15

Gly Leu Asp Asn Gln Tyr Leu Lys Ala Gly Asp
20 25

<210> SEQ ID NO 25
<211> LENGTH: 18
<212> TYPE: PRT

-continued

<213> ORGANISM: *Pseudomonas aeruginosa*

<400> SEQUENCE: 25

Leu Ser Thr Ala Ala Asp Met Gln Gly Val Val Thr Asp Gly Met Ala
1 5 10 15

Ser Gly

<210> SEQ ID NO 26
<211> LENGTH: 53
<212> TYPE: PRT
<213> ORGANISM: *Pseudomonas aeruginosa*

<400> SEQUENCE: 26

Pro Gly Asn Leu Pro Lys Asn Val Met Gly His Asn Trp Val Leu Ser
1 5 10 15

Thr Ala Ala Asp Met Gln Gly Val Val Thr Asp Gly Met Ala Ser Gly
20 25 30

Leu Asp Lys Asp Tyr Leu Lys Pro Asp Asp Ser Arg Val Ile Ala His
35 40 45

Thr Lys Leu Ile Gly
50

- What is claimed is:
1. A pharmaceutical composition, comprising:
at least one peptide that is a truncation of a cupredoxin and
that can inhibit the development of premalignant lesions
in mammalian tissue; and
a pharmaceutically acceptable carrier.

2. The pharmaceutical composition of claim 1 which com-
prises at least two of the peptides.

3. The composition of claim 1, wherein the pharmaceutical
composition is formulated for intravenous administration.

4. The composition of claim 1, wherein the cupredoxin is
from an organism selected from the group consisting of
Pseudomonas aeruginosa, *Alcaligenes faecalis*, *Achromo-
bacter xylosoxidans*, *Bordetella bronchiseptica*, *Methylomo-
nas* sp., *Neisseria meningitidis*, *Neisseria gonorrhea*,
Pseudomonas fluorescens, *Pseudomonas chlororaphis*,
Xylella fastidiosa and *Vibrio parahaemolyticus*.

5. The composition of claim 4, wherein the cupredoxin is
from *Pseudomonas aeruginosa*.

6. The composition of claim 1, wherein the cupredoxin is
selected from the group consisting of SEQ ID NOS: 2 and
20-26.

7. A kit comprising the composition of claim 1 in a vial.

8. The kit of claim 7, wherein the kit is designed for intra-
venous administration.

9. A pharmaceutical composition, comprising:
at least one peptide that is a truncation of a cupredoxin in a
therapeutically effective amount to inhibit the develop-
ment of premalignant lesions in a mammalian tissue;
and
a pharmaceutically acceptable carrier.
- * * * * *